

UCLA

UCLA Previously Published Works

Title

Stress-related alterations of visceral sensation: animal models for irritable bowel syndrome study.

Permalink

<https://escholarship.org/uc/item/23z6778m>

Journal

Journal of neurogastroenterology and motility, 17(3)

ISSN

2093-0879

Authors

Larauche, Muriel
Mulak, Agata
Taché, Yvette

Publication Date

2011-07-01

DOI

10.5056/jnm.2011.17.3.213

Peer reviewed

Stress-Related Alterations of Visceral Sensation: Animal Models for Irritable Bowel Syndrome Study

Muriel Larauche,* Agata Mulak and Yvette Taché

CURE/Digestive Diseases Research Center and Center for Neurobiology of Stress, Digestive Diseases Division, Department of Medicine, David Geffen School of Medicine, UCLA and VA Greater Los Angeles Healthcare System, Los Angeles, California, USA

Stressors of different psychological, physical or immune origin play a critical role in the pathophysiology of irritable bowel syndrome participating in symptoms onset, clinical presentation as well as treatment outcome. Experimental stress models applying a variety of acute and chronic exteroceptive or interoceptive stressors have been developed to target different periods throughout the lifespan of animals to assess the vulnerability, the trigger and perpetuating factors determining stress influence on visceral sensitivity and interactions within the brain-gut axis. Recent evidence points towards adequate construct and face validity of experimental models developed with respect to animals' age, sex, strain differences and specific methodological aspects such as non-invasive monitoring of visceromotor response to colorectal distension as being essential in successful identification and evaluation of novel therapeutic targets aimed at reducing stress-related alterations in visceral sensitivity. Underlying mechanisms of stress-induced modulation of visceral pain involve a combination of peripheral, spinal and supraspinal sensitization based on the nature of the stressors and dysregulation of descending pathways that modulate nociceptive transmission or stress-related analgesic response.

(J Neurogastroenterol Motil 2011;17:213-234)

Key Words

Irritable bowel syndrome; Models, animal; Pain

Introduction

Alterations of visceral sensation such as enhanced perception of physiological or experimental visceral stimuli along with hypervigilance to those, are at the origin of visceral hypersensitivity,

a phenomenon commonly considered to play a major role in the pathophysiology of irritable bowel syndrome (IBS).¹⁻⁷ Epidemiological studies have implicated stress of psychosocial, physical or immune origin as a trigger of first onset or exacerbation of IBS symptoms.⁸⁻¹⁰ Early adverse life events in the form of emotional, sexual, or physical abuse are major predisposing factors for the

Received: June 9, 2011 Revised: None Accepted: June 12, 2011

© This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Correspondence: Muriel Larauche, PhD

CURE/Digestive Diseases Research Center, West Los Angeles VA Medical Center, 11301 Wilshire Blvd., Bldg. 115/Rm 111, Los Angeles, CA 90073, USA

Tel: +1-310-478-3711, Fax: +1-310-268-4963, E-mail: mlarauche@mednet.ucla.edu

Financial support: This review is part of studies supported by the VA Research Career Scientist Award, NIH grants R01 DK-57238 and DK 33061 and P50 DK-64539 (YT) and K01 DK088937 (ML).

Conflicts of interest: None.

development of IBS later in life.^{11,12} Childhood trauma, especially in genetically predisposed individuals, is thought to induce persistent changes in the brain arousal response system that impacts on the neuroendocrine hypothalamic-pituitary-adrenal (HPA) axis.¹² In adult IBS patients, acute stress episodes, chronic social stress, anxiety disorders, and maladaptive coping style determine the illness experience, health care-seeking behavior as well as treatment outcome.^{12,13} Stress-related psychosocial factors such as somatization, neuroticism, and hypochondriasis are also important predictors in the development of post-infectious IBS.^{14,15} Emotional or physical stressors may cause disturbances at every levels of the brain-gut axis including the central, autonomic and enteric nervous systems and affect regulation of visceral perception and emotional response to visceral events.¹⁶

Over the past 15 years, various animal models have been developed to get insight into the underlying mechanisms of visceral hypersensitivity and the influence of stress on visceral pain pathways.^{1,17-20} While in humans the evaluation of visceral sensitivity is predominantly based on the conscious perception to gut distension, the measurement of this subjective response cannot be performed in animal studies. Objective evaluation of responses to visceral stimulation in clinical studies includes the assessment of reflex activity (eg, a somatic nociceptive cutaneo-muscular flexion reflex can be inhibited by painful visceral stimulation) or evoked central processes (eg, changes in activation of the anterior cingulate cortex involved in pain inhibition).^{21,22} Indeed, during the last decade functional imaging techniques have been applied successfully to examine the human brain response to noxious visceral stimuli.²³ In experimental animals, the pattern of brain and spinal circuitries activated by various stressors and colorectal distension (CRD) under basal or hypersensitive state have been early on mapped in a number of studies using the induction of the Fos protein expression as a direct marker of neuronal cell activation and double immunohistochemical labeling to identify the phenotype of Fos positive spinal and supraspinal neurons.²⁴⁻³¹ Recently, preliminary reports applied imaging techniques to get insight into brain circuit activated by visceral stimulation in rodents. Similarity in some regional brain activation induced by CRD have been found when comparing Fos expression and functional magnetic resonance imaging.³² In addition this comparative study indicates that both methods are complementary as Fos immunohistochemistry provides a higher spacial resolution over imaging while imaging displays a higher sensitivity to detect a large number of brain area. Development of imaging in conscious animals with removal of additional stress linked with conditions

of functional imaging monitoring will enable bridging the gap between the multidimensional nature of human pain experience and preclinical studies.³³

In this review we will outline some of the most relevant preclinical models that have been developed, comment on their contribution to our understanding of stress modulation of visceral pain mechanisms, and assess the clinical relevance of these preclinical studies to unravel potential molecular targets to alleviate visceral pain symptoms in IBS.

Stress Pathways: Corticotropin Releasing Factor Signaling as an End Point Effector

First coined by the endocrinologist Hans Selye, the term “stress” defines the physiological adaptive responses to real or perceived emotional or physical threats (“stressors”) to the organism homeostasis.³⁴ When exposed to an acute threatening challenge, the body engages a “fight or flight” response³⁵ driven by sympathetic activation leading to rapid heart rate and respiration, increased arousal, alertness, and inhibition of acutely non adaptive vegetative functions (feeding, digestion, growth and reproduction).³⁴ Concurrently, a negative feedback is activated to terminate the stress response and bring the body back to a state of homeostasis or eustasis,³⁶ that engages neural, neuroendocrine and immune components, a process called allostasis³⁷ or “stability through changes”.^{37,38} However, persistence or chronicity of the stressors can overload this adaptive system which then becomes defective or excessive. The organism is no longer brought back to basal homeostasis leading to a state of allostatic load^{37,39} or “cacostasis”.³⁶ This state lies at the origin of a variety of stress-related diseases that develop in the context of a vulnerable genetic, epigenetic and/or constitutional background.³⁶ The pathogenesis of stress-induced disorders affects the whole body, including the viscera of which the gastrointestinal (GI) tract is a sensitive target.^{36,40}

Over the past decades, important components of the stress-activated pathways whereby the brain translates stimuli into final integrated bodily response have been identified through the characterization of corticotropin releasing factor (CRF) signaling system. This is composed of the 41 amino acid peptide CRF, and related peptides, urocortin 1, urocortin 2 and urocortin 3 along with the CRF receptors CRF₁ and CRF₂ and their variants which display specific affinity for CRF and related agonists.⁴¹ The development of selective CRF receptor antago-

nists has also largely contributed to delineate the role of activation of CRF receptor subtypes in the stress response.^{42,43} In particular convergent reports indicate that the activation of CRF₁ receptor underlies the multiple faceted components of the stress response.^{40,44,45} CRF/CRF₁ signaling plays a primary neuroendocrine role in stimulating the HPA axis leading to the release of adrenocorticotrophic hormone and corticosterone in rodents and cortisol in humans.^{43,46} In addition the CRF signaling system also acts as a neurotransmitter/neuromodulator to coordinate the behavioral, immune, and visceral efferent limbs of the stress response.^{44,45,47-49} It does so via the activation of the locus coeruleus and its noradrenergic projections to the forebrain which contribute to arousal, alertness as well as the modulation of forebrain, hindbrain and spinal sites regulating the autonomic nervous system activity leading to the stimulation of the sympathetic nervous system and release of catecholamines,⁵⁰⁻⁵² and sacral parasympathetic activity while decreasing vagal efferent output⁵³⁻⁵⁵ that influences immune and visceral function.^{56,57} In addition the brain CRF/CRF₁ signaling pathway is involved in stress-related induction of anxiety/depression^{44,45,58} and alterations of colonic motor and visceral pain while both central and peripheral CRF₂ receptor activation may exert a counteracting influence.⁵⁹⁻⁶³ Moreover recent experimental and clinical studies point to an equally important contribution of the peripheral CRF/CRF₁ signaling locally expressed in the gut to the GI stress response.^{19,64}

Visceral Pain Pathways

Pain perception in peripheral tissues depends on the signal transmission from the site of pain origin to the CNS. Nociceptors (receptors activated by noxious stimuli)⁶⁵ located in 2 sets of primary small afferent fibers (C and A δ afferents) innervating the viscera that project to distinct regions in the CNS,⁶⁶ are the primary pathways of pain transmission. From the esophagus to the transverse colon, the GI tract innervation is provided by vagal afferent fibers originating in the nodose ganglia and projecting centrally to the nucleus of the solitary tract. Pelvic nerve afferent fibers, which originate in the lumbosacral dorsal root ganglia, and project centrally to the lumbar 6 - sacral segments of the spinal cord innervate the remaining part of the large bowel (descending and sigmoid colon, rectum). The entire GI tract is also innervated by afferent fibers contained in the splanchnic nerves projecting to the thoracic 5 - lumbar 2 segments of the spinal cord.⁶⁷ Even though visceral afferents constitute only 10% of all afferents, they are able to monitor changes in the gut milieu and

participate in the transmission of visceral sensory information.^{68,69} Of note, vagal afferents do not encode painful stimuli however, changes in their activity can modulate nociceptive processing in the spinal cord and the brain.^{68,70,71}

Upon entering the dorsal horn, visceral primary afferents carried out by the pelvic and splanchnic nerves terminate in spinal cord laminae I, II, V and X⁷² converges onto spinal neurons in the lumbosacral segments and thoracolumbar segments respectively. Lumbosacral segments process reflex responses to acute visceral pain, while thoracolumbar segments' involvement in normal visceral sensation is uncertain,⁷³ however, both segments process inflammatory stimuli.⁷³ Subpopulations of neurons within the dorsal horn project to discrete nuclei within the thalamus (ie, ventral posterior lateral thalamus) as well as other structures in the brain stem (parabrachial nucleus, periaqueductal gray, nucleus tractus solitarius). From the thalamus, the information is conveyed to cortical areas involved in sensory processing (such as the somatosensory cortex) or those involved in processing emotional or affective information (such as the anterior cingulate gyrus and insular cortex).^{65,74}

In addition to the ascending system, which enables pain perception described above, other neural circuits originating from supraspinal sites can influence nociceptive activity in the spinal cord and in primary afferents, a system referred to as descending pathways.⁷⁵ There are 2 types of descending control pathways: inhibitory, which produce analgesia (periaqueductal gray, locus coeruleus) and facilitatory which produce hyperalgesia (rostromedullary medulla and OFF and ON cells).^{76,77}

Visceral Pain Monitoring in Rodents

The primary readout and the standard assay for the measurement of visceral pain in rodents consists in the monitoring of abdominal muscles contraction or visceromotor response (VMR) to controlled isobaric distensions of the distal colon by an inflatable balloon.⁷⁸ The VMR can directly be assessed as electromyographic (EMG) signals monitored via surgically-implanted recording electrodes in external or internal abdominal muscle which are either externalized through the skin (abdomen, neck)⁷⁹⁻⁸¹ or connected to radiotelemetric implants in the abdominal cavity.^{82,83} Although the method is of significant value in the field of visceral pain study, it has experimental shortfalls such as damage to EMG electrodes, loss of signal and electrical interferences which is of particular concerns in chronic experimental settings. Additionally, EMG surgery involves skin and/or mus-

cle incision depending on the technique used (subcutaneous abdominal electrodes or intraperitoneal cannula) and chronic implantation of a foreign body. Even though no data are available in the literature in relation to the impact of chronic EMG electrodes placed into the abdominal wall, such intervention could induce a host-tissue response with local micro-inflammation (neutrophils, lymphocytes and macrophages) as it has been shown for other types of implants in the skin and peritoneum.^{84,85} A recent report

suggests that the preconditions of animals (EMG surgery, and post-surgical delivery of antibiotic and single housing) has considerable impact on their visceral pain responses, particularly in the context of stress studies.⁸⁶ Other approaches consist of recording manometric changes in the pressure of the balloon inserted into the distal colon^{86,87} or changes in pressure inside the colonic lumen.^{19,88} These 2 later techniques present the advantage of being minimally invasive as they do not require surgery and

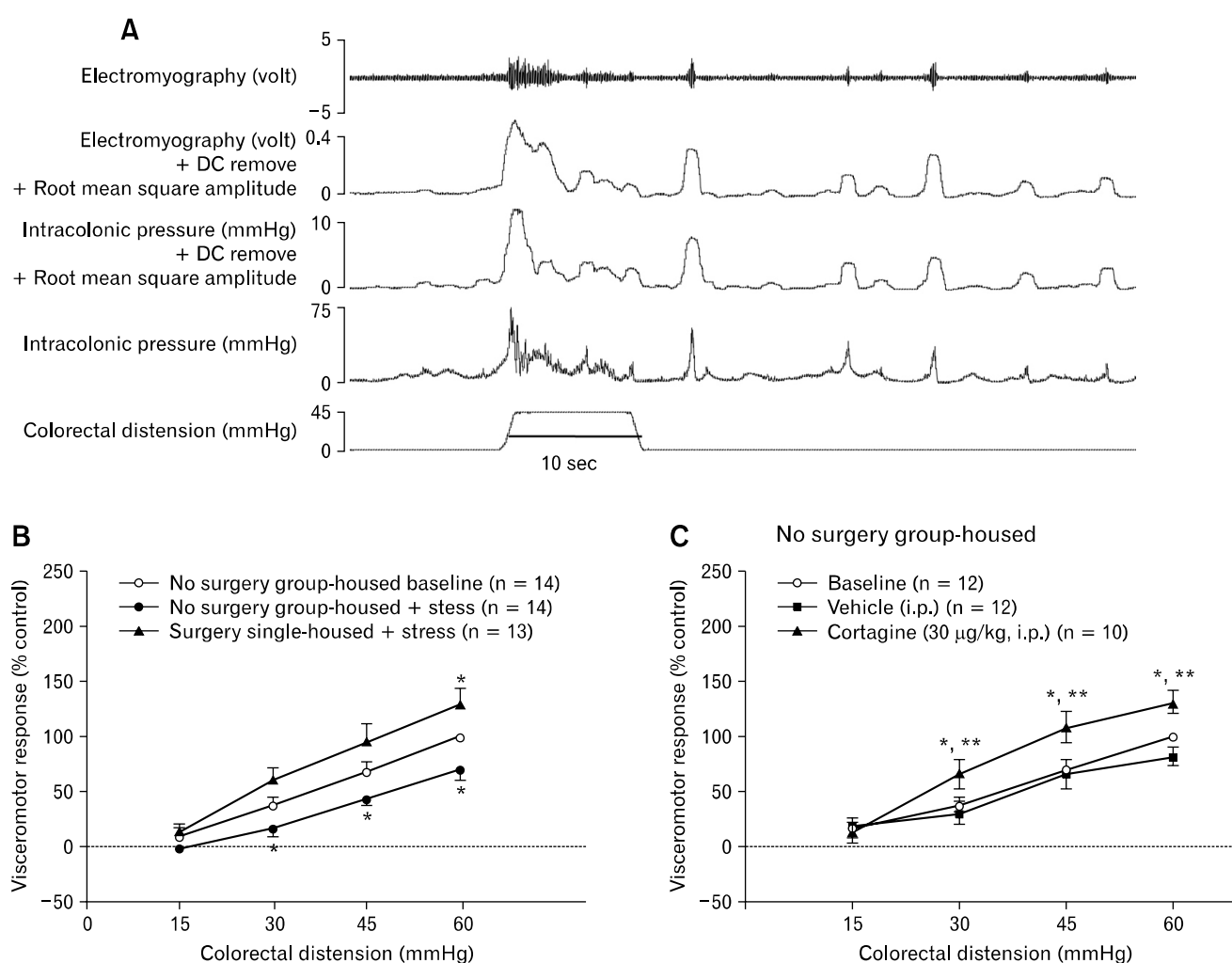


Figure 1. Differential influence of intermittent repeated stress on visceral response to colorectal distension (CRD) in rodents with or without surgical procedure for recording visceral pain (Adapted from Larauche et al^{19,88}). (A) Original and rectified representative electromyographic (EMG) and intraluminal colonic pressure (ICP) traces recorded simultaneously on the same mouse in response to CRD (45 mmHg, 10 seconds). When both raw EMG (upper line) and ICP (second line to the bottom) signals are analyzed in Spike 2 by computing “DC Remove” 1 second to exclude all slow events > 2 seconds (ie, colonic smooth muscle contractions) and “root mean square amplitude” to extract the area under the curve of the signal, the resulting EMG and phasic ICP signals (middle lines) present a significant overlap. (B) Mice were equipped with EMG electrodes or not and exposed to water avoidance stress for 1 hour per day for 10 days tested with ICP for visceromotor response (VMR) to CRD. (C) Intraperitoneal injection of the selective corticotropin releasing factor receptor subtype 1 agonist, cortagine-induced visceral hypersensitivity in C57BL/6 mice tested with ICP for VMR to CRD. Data are expressed as mean \pm SEM, n = 10–14 per group as specified in graph legends. * P < 0.05 compared with baseline, ** P < 0.05 vs vehicle.

post-surgical treatments such as antibiotic, analgesics which can affect the visceral pain responses and still remain an objective and sensitive measure of abdominal contractions (Fig. 1). However, they require the animals to be partially restrained in Bollman cages, a context to which they need to be habituated and which by itself may bring a component of stress.

Behavioral approaches such as operant behavioral assays⁷⁸ have also been used in early studies and capitalized on the learning and fear behaviors of animals in response to painful CRD. Visual monitoring of the abdominal withdrawal reflex⁸⁹ has also been applied in a few studies, and while having the great advantage

of being one of the less invasive technique employed to date, it is a very subjective method. Indirect endpoints such as Fos or extracellular signal-regulated protein kinase induction in the CNS,^{29,62,90-92} and functional brain imaging of integrated brain responses to nociceptive stimuli^{33,93} have also been utilized in some studies. These approaches allow for direct assessment of the neuronal circuitries recruited by the visceral pain stimulus and, in the case of functional brain imaging is very similar to the monitoring of CRD responses in healthy and IBS human subjects. Unfortunately, in animals these brain mapping techniques require euthanasia and limit the assessment to specific time points.

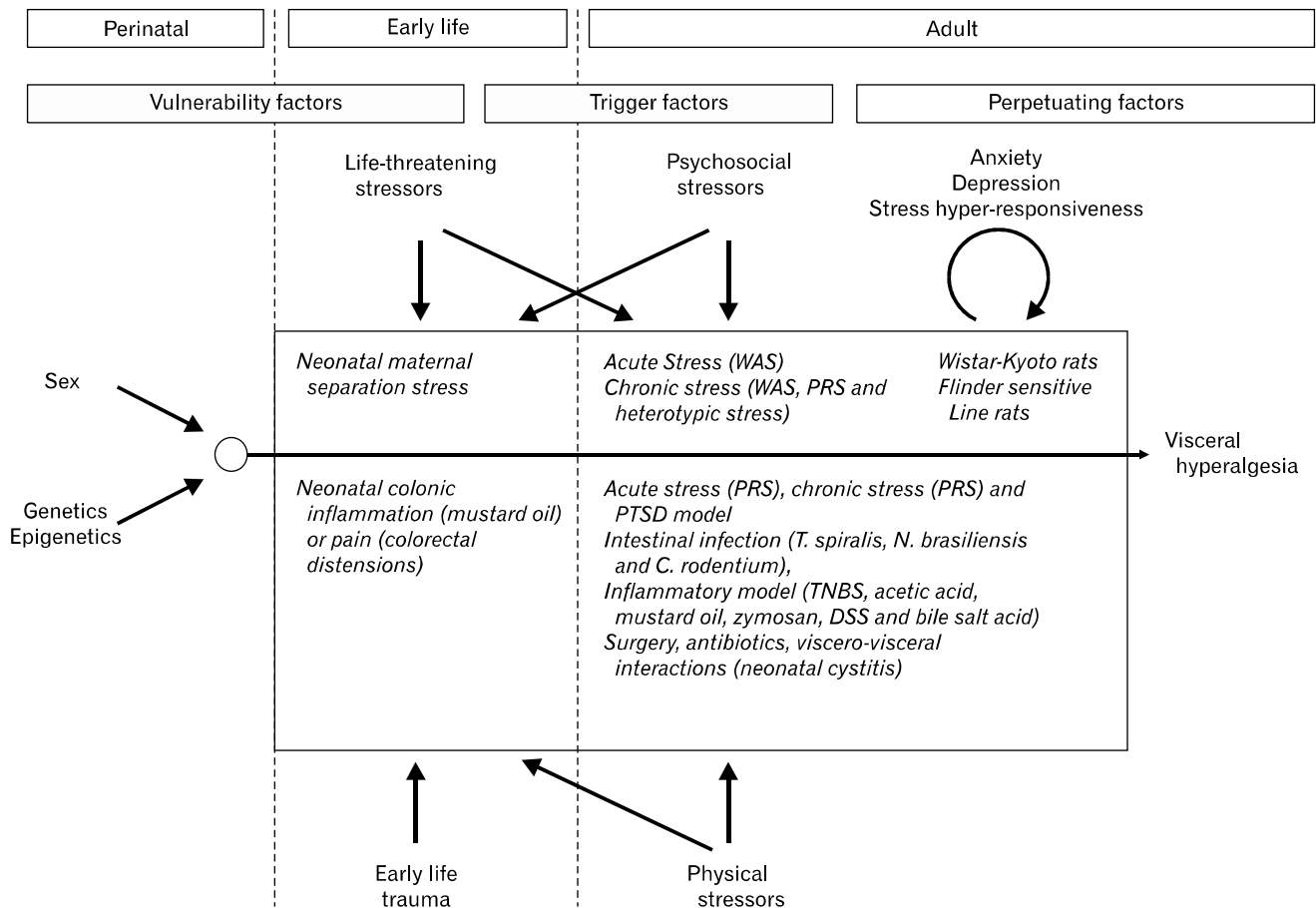


Figure 2. Animal models of stress-induced modulation of visceral sensitivity throughout the lifespan (Modified from Mayer et al⁹). Experimental stress models have been developed that target different periods throughout the lifespan of animals to assess the vulnerability, trigger and perpetuation influences of stress on visceral sensitivity. During early life, trauma due to maternal neglect (neonatal maternal separation stress) or injury (neonatal chronic colonic inflammation or pain) can enhance the susceptibility of individuals to develop altered visceral pain responses at adulthood. During adulthood, life-threatening stressors (post-traumatic stress disorder model), psychosocial stressors (acute and chronic stress) or physical stressors (intestinal infection or inflammation, antibiotic administration and surgery) have all clearly been established as triggering factors to the development of visceral hypersensitivity in rats and mice. Lastly, the use of specific strains of rodents known to exhibit various levels of anxiety, depression or stress hyper-responsiveness (Wistar-Kyoto and Flinders Sensitive Line) help mimic the influence of perpetuating factors on symptoms of visceral pain. WAS, water avoidance stress; PRS, partial restraint stress; PTSD, post-traumatic stress disorder; DSS, dextran sodium sulfate.

However, as more stringent brain imaging approaches are developed in rodents, they will open new venues to parallel human studies.⁹⁴

Experimental Stress Models and Visceral Pain

By convention, stressors are categorized in exteroceptive (psychological or neurogenic) and interoceptive (physical or systemic) classes^{95,96} and both have been used in animal models to investigate the relationship between stress and visceral pain.⁹⁷ Dual visceral pain responses: hyperalgesia and analgesia have been described in rodents exposed to exteroceptive stressors. Though only recently investigated, the analgesic response bears very relevant implications in the understanding of visceral pain-associated pathologies (detailed in section “Stress-induced visceral analgesia: how does it help us to model and understand visceral hypersensitivity?”) In contrast, interoceptive stressors have been most consistently associated with the development of stress-induced hyperalgesia.

Stress modulates visceral pain in IBS patients as well as in healthy subjects,^{9,98} therefore experimental animal models, involving exposure to various clinically relevant stressors have been developed to recapture features of IBS symptoms, of which hyperalgesia to sigmoid distensions is a hallmark.^{99,100} Moreover clinical studies have stratified that the stress-related modulation of IBS symptoms⁹ may be occurring through (1) permanent enhancement of stress responsiveness, (2) transient symptom exacerbation and/or (3) symptom perpetuation. Consequently existing experimental stress models target different periods throughout the lifespan of animals to assess the vulnerability, the trigger and perpetuating factors determining stress influence on visceral hypersensitivity (Fig. 2).

Stress in the Perinatal Period: Genetic/Epigenetic Factors

Twin studies in IBS patients showed higher (but relatively low) concordance rates in monozygotic than dizygotic twins suggesting that although genetic factors are not dominant, they play a role in the occurrence of IBS.¹⁰¹ There is also a growing literature reporting the association between functional genetic polymorphisms and IBS at the level of serotonin transporter gene (associated with diarrhea in female IBS patients), or α_2 -adrenoreceptor gene (associated with constipation), and more recently, additional gene polymorphisms have been unraveled supporting

the potential permissive role of genetics in IBS pathophysiology.¹⁰²⁻¹⁰⁵ Of interest, it has been postulated that epigenetic factors related to heritable changes in gene expression that occur without alteration in gene sequence, determine the manner in which gene activity may be altered either transiently or permanently in response to environmental challenges.¹⁰⁶ Such epigenetic modifications could account for symptoms persistence, familial clustering and the transgenerational impact of IBS. However, experimental studies have not dwelled on strain differences in terms of stress responsiveness, anxiety and depression in rodents,¹⁰⁷⁻¹¹⁰ to assess and compare how genetic predisposition together with perinatal (maternal prenatal stress) or early life stressors (neonatal maternal separation stress) could affect visceral pain responses at adulthood in the context of epigenetic modifications. There is only one preliminary report suggesting that strain may determine the duration of the visceral hyperalgesia in response to chronic heterotypic stress (detailed in section “Stress in the adult period: trigger factors, Psychosocial stressors”). The influence of genes on the vulnerability of rodents to exhibit visceral hypersensitivity has however been assessed in relation with anxiety behavior at adulthood in rat strains with different anxiety/depression backgrounds¹¹⁰ (detailed in section “Genetic models of anxiety & depression”).

Stress in the Early-life Period: Vulnerability/Trigger Factors

Early life events and childhood trauma by biopsychosocial factors (neglect, abuse, loss of caregiver or life threatening situation) enhance the vulnerability of individuals later in life to develop affective disorders (depression, anxiety and emotional distress) and put them at a greater risk for developing IBS.^{12,99} In the context of epigenetic modifications, experimental studies showed that early developmental trauma decreases glucocorticoid receptor expression by hypermethylation of a key regulatory component and consequently affects the feedback regulation the HPA-axis with impact on the endocrine/behavioral adaptation and the susceptibility to stress-related disorders.¹¹² In addition, experimental studies indicate that the newborn's gut through stress-related changes in intestinal permeability may be exposed to a variety of factors resulting in mucosal inflammation and tissue irritation which could have long-term consequences at adulthood even though no longitudinal clinical studies exist showing that gut irritation in early life is a risk factor for IBS development at adulthood.⁹⁷ Moreover, postnatal microbial colonization has been also suggested as a potential factor programming the

HPA-axis response to stress in mice.¹¹³

An experimental model commonly used as a stress model to mimic early stress/childhood trauma is the neonatal maternal separation in rodents. This is achieved by isolating pups from the dam for 2-3 hours/day during the first 2 weeks after birth from postnatal day (PND) 1-2 to PND 14.^{17,114-116} At adulthood, rats previously subjected to neonatal maternal separation exhibit visceral hypersensitivity to CRD under basal conditions which is further exacerbated by exposure to the acute psychological stressor in the form of water avoidance stress (WAS) consisting in placing rodents on a small platform surrounded by water for 1h.^{117,118} Other models used repeated intermittent colonic irritation during the neonatal period (PND 8-21) either in the form of daily noxious CRD (60 mmHg-60 seconds distension twice separated by 30-minute period of rest) or by performing daily intracolonic injection of mustard oil (5%), increases pain behavior to CRD from postnatal week 5 up to postnatal week 12.^{89,119} Likewise, an acute somatic injury (saline or carrageenan injections into the hind paw) performed during the critical period of postnatal development, ie, before PND 14, produces visceral analgesia to CRD in adult rats.¹²⁰

Based on these studies and the extensive amount of evidence originating from somatic pain studies,^{121,122} it appears that neonatal insults either acute or repeated, somatic vs visceral occurring during the development of the organism contribute to induce a state of visceral hypersensitivity in adulthood which may reflect long-term changes in visceral sensory processing.¹²⁰

Stress in the Adult Period: Trigger Factors

Psychosocial stressors

Psychosocial stressors (eg, threat to social status, social esteem, respect and/or acceptance within a group; threat to self-worth) activate stress circuits within the emotional motor system and induce neuroendocrine response (CRF and cortisol) and autonomic response (norepinephrine and epinephrine) that result in the modulation of gut sensory, motor and immune function as well as intestinal permeability.⁹ In experimental studies, the 2 main acute stressors that are prominently used in visceral pain studies are WAS for 1 hour and partial restraint stress for 2 hours, a stressor with stronger psychological component than WAS, which entails taping the forelimb of rats in order to prevent their movements.¹²³⁻¹²⁵ Exposure of male Wistar rats to WAS for 1 hour leads to the development of a delayed visceral hyperalgesia to CRD, appearing 24 hours after the end of the stress,¹²⁶ while exposure to partial restraint stress, induces an immediate hyper-

algesia to CRD in male¹²⁷ and female Wistar rats.¹¹⁵

However, in the context of clinical studies in which daily chronic stress predicts the intensity and severity of subsequent symptoms in IBS patients,^{4-6,99,128,129} a variety of chronic stress models divided in 2 categories have been recently developed in adult rodents. The first category consists in exposing animals repeatedly (over a few days to weeks) but intermittently (once or twice per day) to 1 or different stressors, with the aim of mimicking the daily exposure to psychosocial stress that humans can encounter through their personal and professional interactions. The second category consists in continuous exposure to stressors as part of change in internal state, for instance inflammation, or external milieu, for instance single housing, or social crowding which mimics the effect of social milieu in humans or using genetic rodent strains that have constitutive stress hyper-reactivity (Wistar Kyoto, Flinders Sensitive Line). In particular, repeated intermittent exposure to WAS is one of the first "chronic" stress model to have been adapted to the study of visceral hypersensitivity⁸¹ and is presently widely used.^{88,130,131} Initial studies in which the visceral pain response was monitored using EMG recording that entails surgical implantation of electrodes, male Wistar rats exposed to 10 consecutive days of WAS for 1 hour daily developed visceral hypersensitivity to CRD lasting up to 30 days after the end of the last session of WAS.^{81,130} In our laboratories however, we found that when naïve male and female Wistar rats were exposed to a similar WAS schedule and their VMR was monitored by intraluminal colonic solid-state manometry, a technique that does not require surgery, animals developed visceral analgesia to CRD.¹³² Similar results have been obtained in C57BL/6 mice⁸⁸ and analgesic vs hyperalgesic responses were established to be dependent upon preconditions (surgery and single housing) associated with the method of recording of VMR (Fig. 1).^{88,133} Therefore, the impact of repeated mild stress such as 1-hour exposure to WAS on visceral pain response to CRD is largely influenced by the basal state conditions of the animals before applying the repeated stressor (detailed in section "Stress-induced visceral analgesia: how does it help us to model and understand visceral hypersensitivity?" and reference⁸⁸). Repeated exposure to unpredictable sound stress has also been recently shown to provide a model of delayed visceral hyperalgesia in male Sprague-Dawley rats.¹³⁴

Because habituation can occur in response to repeated exposure to an homotypic stressor,^{135,136} heterotypic stress models with different and alternating modalities to induce stress have been recently developed. However male Wistar rats exposed ran-

domly to a combination of cold restraint stress (45 minutes), WAS (1 hour) or forced swimming (20 minutes), 1 stressor per day for 9 consecutive days develop visceral hypersensitivity only at 8 hours but not at 24 hours or 7 days after the end of the last stressor.¹³⁷ Interestingly however, the same regimen of alternating stressors in a different strain of rats, Sprague-Dawley, led to a sustained visceral hypersensitivity lasting up to 2 weeks after the end of the stressor (S. Sarna and J. Winston, pers. comm.), suggesting that the strain and therefore genetic background of the animals, affects the visceral pain responses to repeated intermittent exposure to different stressors.

Life-threatening stressors

Retrospective clinical studies indicate that living through or seeing a traumatic event, such as war, environmental disasters, rape, physical abuse or a bad accident in adulthood can lead to post-traumatic stress disorder (PTSD).¹³⁸⁻¹⁴⁴ There is evidence of increased prevalence of GI symptoms, in particular IBS in PTSD sufferers including war veterans.¹³⁸⁻¹⁴² Additionally, patients with IBS who have experienced traumatic events may be at higher risk for other co-morbid psychiatric disorders than IBS patients without a trauma history.¹⁴¹

In adult rats, treatment with a relatively short-lasting session of shocks or a social confrontation with a predator or aggressive conspecific animals induces long-lasting (weeks-months) conditioned fear-responses to trauma-related cues, and a generalized behavioral sensitization to novel stressful stimuli that persists or grows stronger over time.¹⁴⁵⁻¹⁴⁸ Repetitive balloon distention of the distal colon causes increased cardiovascular 'pseudoaffective' reflexes in pre-shocked rats compared to controls, 2 weeks after a single session of foot shocks.¹⁴⁵⁻¹⁴⁸ Of note, female rats appear to show a different pattern of sensitized behavioral responsiveness to the same challenge, possibly pointing to sex-related alterations in the neuronal substrates involved.¹⁴⁹

Interoceptive stressors

In approximately 10% of patients with IBS, the onset of symptoms began with an intestinal infectious illness.¹⁵⁰ Bile salt malabsorption resulting from infectious damage with organisms such as *Salmonella* and *Campylobacter* within the terminal ileum and right colon may also underlie some forms of post-infectious IBS.¹⁵¹ Inflammation, antibiotic treatments, bladder infection and surgery may also contribute to the symptoms in some patients. Below are described some experimental models of interoceptive stressors that have been used to mimic these clinical conditions.

Post-infectious irritable bowel syndrome model. Pro-

spective studies have shown that 3% to 36% of enteric infections lead to persistent new IBS symptoms depending on the infecting organism. In addition, the co-existence of adverse psychological factors at time of infection is also an important determinant to the susceptibility to develop post-infectious IBS.¹⁵² Other risk factors include female sex and some psychological characteristics such as anxiety, depression and somatization.¹⁵² While viral gastroenteritis seems to have only short-term effects, bacterial enteritis and protozoan and helminth infestations are followed by prolonged post-infectious IBS.¹⁵² The vast majority of human post-inflammatory hypersensitivity symptoms are observed after bacterial infection (*Campylobacter*, *Shigella*, *Salmonella* or *Escherichia coli* infections).

In preclinical models, long-lasting visceral hyperalgesia has been observed in mice after transient intestinal inflammation induced by *Trichinella spiralis* infestation^{153,154} or in rats infested by *Nippostrongylus brasiliensis*.¹⁵⁵ Recently, however, it was found that male C57BL/6 mice infected with *Citrobacter rodentium*, an attaching-effacing murine enteropathogen similar in its mechanisms of infection to enteropathogenic *Escherichia coli*, do not spontaneously develop visceral hypersensitivity symptoms assessed by the increase in EMG response to CRD¹⁵⁶ unless exposed to a stressor (WAS, 1 hr/day for 9 days) during the time of infection (S. Vanner and N. Cenac, pers. comm.).

Post-inflammatory irritable bowel syndrome model.

Despite some controversies on the origin of the symptoms,^{157,158} "IBS-like" symptoms appear to be common in patients in remission from ulcerative colitis.^{15,159} In rats, chemical irritants applied to the colon such as acetic acid,¹⁶⁰ mustard oil^{161,162} and zymosan^{163,164} evoke short-term hyperalgesia associated with transmural tissue damage/colonic inflammation. Intracolonic trinitrobenzene sulfonic acid induces a severe colonic transmural inflammation and visceral hypersensitivity that develops at 4-5 days with the disappearance of symptoms by 14 days.^{165,166} Interestingly, in 24% of rats there is reoccurrence of visceral hyperalgesia 16 weeks after the induction of inflammation, while there is no evidence of microscopic inflammation in rat colonic tissues at this time point.^{166,167} In a similar manner, daily intracolonic instillation of bile acid deoxycholic acid for 3 days induces a mild, transient colonic inflammation within 3 days of administration that resolves within 3 weeks in adult male Sprague-Dawley rats. In this model, a persistent visceral hyperalgesia starts after 1 week of bile acid administration which lasts up to 4 weeks.¹⁶⁸

Mild non-specific colitis and acute dextran sodium sulfate (DSS, 5% in drinking water for 5 days)-induced colitis have been

associated with increased responsiveness to CRD on days 5 or 60 after the induction of colitis in male Balb/c mice while chronic colitis induced by DSS (3 cycles of 5% DSS for 5 days/cycle and 15 days of normal drinking water in between each cycle) has not.¹⁶⁷ These results are in contrast with another study showing that 4% DSS in drinking water for 5-7 days-induced colitis but failed to cause the development of visceral hypersensitivity in response to CRD in C57BL/6 or Balb/c mice when tested on days 5, 12, 16, 20, 30, 40 or 51 after the induction of colitis.¹⁷⁰ These disparate findings suggest that inflammation alone may not always lead to visceral hypersensitivity and that the type of inflammatory insult and severity determine whether this will result in the development of postinflammatory hypersensitivity. The interaction between colonic inflammation and the development of visceral pain has to be substantiated in future investigations.¹⁶⁶

Antibiotics. Patients treated with antibiotics for non-GI complaints are 3 times more likely to report functional bowel symptoms. Antibiotic use disrupts the intestinal microbiota, fragilizes the host's intestinal homeostasis and integrity of intestinal defenses,¹⁷¹ and has been associated with IBS.¹⁷² In support of this hypothesis, administration to Balb/c mice of an oral combination of non-absorbable antibiotics (neomycin, bacitracin and pimaricin) which disturbed the commensal intestinal microflora results in visceral hypersensitivity to CRD in these animals.¹⁷³ Paradoxically, clinical studies support that specific antibiotics (rifaximin or neomycin) are an effective treatment option in non-constipated IBS patients, over a 3-month period^{174,175} or even longer,¹⁷⁶ thereby confirming the role of dysbiosis in developing IBS symptoms.¹⁷⁷

Surgery and somato-visceral convergence. Despite controversies, studies suggest that IBS is associated with an increased risk of abdominal and pelvic surgeries.¹⁷⁸ Surgical procedure as both a visceral and psychological stressor can initiate a series of events that either disturb GI function and interactions within the brain-gut axis and/or alter gut microbiota, which consequently may lead to generation of IBS symptoms.¹⁷⁹ Hind paw (plantar) incision or injection of low pH (4.0) sterile saline in the gastrocnemius muscle of adult male Sprague-Dawley rats induce a significant visceral hyperalgesia to CRD that lasts up to 2 weeks after the somatic injury occurred.^{180,181} As a model of postsurgical pain, the plantar incision model is particularly relevant because surgical procedures are relatively common and possible visceral hypersensitivity may also thus be a relatively common postsurgical event.¹⁷⁹ The impact of somato-visceral convergence has to be considered in experimental models of visceral pain where

animals are surgically equipped within the abdominal wall with EMG electrodes⁸⁴ (detailed in section "Stress-induced visceral analgesia: how does it help us to model and understand visceral hypersensitivity?").

Viscero-visceral interactions: neonatal cystitis. A significant overlap is observed between IBS and urinary symptoms, in particular those resulting from interstitial cystitis (IC).¹⁸² Like IBS, IC predominantly affects female patients (90%) and shows a high comorbidity rate with psychological disorders. By analogy to IBS, an increased number of mast cells have been found in bladder biopsies in IC.¹⁸³ Recurrent urinary tract infections during childhood correlate with the development of chronic pelvic pain, a condition that often overlaps with IBS.¹⁸⁴ In an animal model of bowel-bladder cross-sensitization, acute bladder chemical irritation causes a significant decrease in colorectal sensory thresholds to CRD.¹⁸⁵ Very recently, the induction of neonatal cystitis in female Sprague-Dawley rats at PND 14 was shown to result in colonic hypersensitivity to CRD during adulthood,¹⁸⁶ supporting a potential key role for viscero-visceral convergence in IBS and comorbid disorders such as IC and chronic pelvic pain.¹⁸²

Stress in the Adult Period: Perpetuating Factors

There is a strong overlap between IBS and psychiatric disorders, as established by the high percentage (54% to even 94%) of IBS patients meeting the criteria for at least 1 primary psychiatric disorder, most notably mood and anxiety disorders.¹⁸² Although comorbid psychiatric disorders seem to be not directly connected with the occurrence of IBS, they strongly influence how the symptoms are experienced, the individual illness behavior, and ultimately the outcome. The influence of cognitive aspects as well as motivational and emotional components on the processing of sensory information is mediated by extensive neuro-anatomical network with a pivotal role of the insular and anterior cingulate cortices.^{9,187,188} Autonomic dysfunction, in particular decreased parasympathetic activity and increased sympathetic outflow observed in psychiatric disorders as well as in IBS,^{16,189,190} has been also suggested to have a relevant impact on the neurally mediated regulation of colonic sensory-motor and immune function.¹⁹¹ The neuroimmune cross-talk involving the stress-induced changes in vagal nerve activity and/or sensitization of mast-cells seems to play a critical role in altering visceral sensitivity and intestinal barrier.¹⁹²

Genetic models of anxiety and depression

In a comparative study using 3 strains of rats known to have

varying levels of baseline anxiety, the high-anxiety Wistar-Kyoto rats had increased VMR to CRD compared to low-anxiety Sprague-Dawley and Fisher-344 animals suggesting a direct link between anxiety and visceral hypersensitivity.¹¹¹ In addition, compared to low-anxiety strains of rats, the sensitivity of high-anxiety rats was highly exacerbated by peripheral sensitization of the colon with a small dose of acetic acid.¹¹¹ Of note, Wistar-Kyoto rats are also considered as a model of depression,^{193,194} as are rats from the Flinders Sensitive Line which exhibit increased cholinergic sensitivity compared to control rats of the Flinders Resistant Line.^{195,196} Similarly to Wistar-Kyoto rats, Flinders Sensitive Line rats exhibit increased VMR to CRD as well as a blunted corticosterone response to acute noise stress compared to Flinders Resistant Line, suggesting a link between depression, HPA axis dysfunction and visceral hyperalgesia.¹⁹⁷

Genetic models of chronic stress

Genetic models that blocked chronically the stress pathways by deleting CRF₁ receptors showed a decrease in anxiety and colonic sensitivity to CRD.¹⁹⁸ Conversely, genetic models of chronic stress relying on the over-expression of CRF stress system in mice¹⁹⁹ are available and could be useful to study IBS-like manifestations, but the visceral sensitivity of these transgenic animals has not been assessed yet. However, as CRF over-expressing mice display phenotypes of Cushing's syndrome,²⁰⁰ new promising genetic models with more selective conditional and/or region-targeted genetic manipulations including RNAi gene silencing technology to modify CRF-related genes are continuously developed.²⁰¹⁻²⁰⁶ These models will be suitable to explore specific stress circuitries in the context of targeted chronic CRF expression/deletion and the impact on visceral pain modulation which so far is lagging behind.

Stress-Induced Visceral Analgesia: How Does It Help Us to Model and Understand Visceral Hypersensitivity? —

While extensively described in somatic pain field,²⁰⁷ to date activation of descending inhibitory pathways in stress-related visceral responses has received less attention. Opioids have been implicated in descending inhibition of visceral sensitivity following an acute stress as evidenced by the fact that naloxone unmasked WAS-induced hyperalgesia to CRD in normal Long-Evans rats and exacerbated the pain response to CRD in maternally-separated rats.¹¹⁷ In another study, a non-opioid, neurotensin-dependent visceral analgesic response was observed 6 hours after ex-

posure to an acute session of WAS in Sprague-Dawley rats, with males exhibiting stronger analgesia than females as well as in wild-type but not in neurotensin knock-out mice.²⁰⁸ In another experimental model, a daily short period (15 minutes) of separation from PND 2 to 14, decreased VMR to CRD performed immediately after WAS and prevented the development of hyperalgesia 24 hours after WAS in adult male Long-Evans rats.²⁰⁹ These data suggest a potential upregulation of endogenous pain-modulatory systems by this short maternal separation stress.²⁰⁹ Similar findings in adult rats have been recently reported, such that Wistar rats handled daily for 9 days develop visceral hypoalgesia in response to CRD that becomes significant 7 days after the last handling.¹³⁷

These studies point to the type of stress itself contributing to the differential recruitment of those descending inhibitory pathways. However, importantly, we recently demonstrated that mice that had undergone surgery for the placement of EMG electrodes on abdominal wall and were subsequently single housed to avoid deterioration of implanted electrodes by cage-mate, developed visceral hyperalgesia in response to repeated WAS (1 hr/day, 10 days) while mice tested for visceral pain using the non-invasive solid-state intraluminal pressure recording and kept group housed developed a strong visceral analgesia under otherwise similar conditions of repeated intermittent WAS.⁸⁸ As mentioned before surgery *per se* is known to induce a long lasting visceral hyperalgesia and recent reports suggest that previous injury or exposure to opioids in male rats can switch stress influence on pain responses from analgesia to hyperalgesia.²¹⁰ Collectively these data demonstrate that the state of the animal tested (naïve vs exposed to surgery), its social environment (group housing vs single housing, cage enrichment or not), the handling performed by the investigator, the methods used to record VMRs (EMG requiring surgery and antibiotic post surgery vs manometry not requiring surgery/antibiotic), as well as the sex of animals can significantly affect the response to exteroceptive stressors. Therefore these preconditions should be carefully detailed in describing the experimental conditions and taken into consideration in the design, conduct and interpretations of the data when investigating the influence of stress on visceral sensitivity in experimental animals.

Based on recent clinical findings demonstrating that IBS patients have compromised engagement of the inhibitory descending pain modulation systems,^{21,211,214} gaining a deeper understanding of the mechanisms involved in the expression of stress-induced visceral analgesia or lack thereof are promising

avenues to be explored and may lead to new therapeutic targets for IBS. Therefore the use of non-invasive methods of monitoring VMR that alleviates the surgical, antibiotic and housing impacts on repeated stress modulation of visceral pain represents a step forward to gain insight into the underlying mechanisms in particular the neural substrates and neurochemistry of stress-related analgesia as established in the somatic field.²⁰⁷

Sex Differences in Stress-Induced Alterations of Visceral Sensitivity

Women are more susceptible to stress-related disorders which is consistent with female predominance in IBS patients (women to men ratio about 2:1).^{215,216} Sex differences in the stress response and stress-induced pain modulation have been documented in a number of human studies.²¹⁷ Clinical trials have also revealed important sex-related differences in therapeutic efficacy of some serotonergic drugs used in IBS treatment (eg, alosetron, 5-HT₃ receptor antagonist) suggesting a conceivable link between estrogens and serotonergic mechanisms in the modulation of stress-related visceral hypersensitivity.^{218,219} Contrasting with this clinical evidence, most of the preclinical studies assessing stress-related alterations in visceral sensitivity have been conducted in male rodents.^{208,220} However, the few studies performed in female indicate that sex hormones have a significant effect on visceral sensitivity in rodents.²²⁰⁻²²⁴ Therefore, addressing the influence of sex and sex hormones in the modulation of visceral pain by stress appears critical to develop novel therapies relevant to sex difference in IBS.^{216,225}

Mechanisms Involved in Stress-Induced Modulation of Visceral Pain

Maladaptive neuroplastic changes leading to persistent increased perception and responsiveness to noxious stimuli, or response to normally non-noxious stimuli are key for the expression of pathological pain (hyperalgesia and allodynia). Such neuroplastic changes can occur in primary afferent terminals (peripheral sensitization) but also in the spinal cord (central sensitization) and in the brain (supraspinal pain modulation) or in descending pathways that modulate spinal nociceptive transmission. Such alterations in the processing of sensory information are all considered as possible mechanisms of visceral hypersensitivity in IBS patients.^{66,226}

Peripheral Sensitization: Corticotropin Releasing Factor System, Mast Cells, Gut Microbiota and Ion Channels

Several reports in both humans and rodents have well documented the key role played by the peripheral CRF signaling, via CRF₁ receptors, in the development and expression of visceral pain.^{19,60,227-231} Stress and peripheral administration of CRF induce mast cells degranulation in the colon in experimental animals and humans,^{232,233} which contributes to the development of visceral hypersensitivity (Fig. 1) via the release of several preformed or newly generated mediators^{118,234-237} (histamine, tryptase, prostaglandin E₂, nerve growth factor) that can stimulate or sensitize sensory afferents^{66,238} by activating a number of ion channels widely expressed in colonic afferents²³⁹⁻²⁴² such as N-methyl-D-aspartate receptor,²⁴² proteinase-activated receptor,²³⁶ and transient receptor potential vanilloid 1²⁴³⁻²⁴⁵ to name a few.

Stress can also disrupt the intestinal epithelial barrier thereby increasing the penetration of soluble factors (antigens) into the lamina propria, leading to nociceptors sensitization,^{235,246} a phenomenon which appears as a prerequisite for the development of visceral hypersensitivity in both humans and rodents.²⁴⁶⁻²⁴⁸ Alterations of epithelial permeability following stress involves the activation of the peripheral CRF system and may²⁴⁹⁻²⁵³ or may not be dependent from mast cell activation^{238,253} in a time-dependent manner. In addition to inducing a leaky epithelial barrier, stress can also change the composition of the intestinal and fecal microbiota of rodents.²⁵⁴⁻²⁵⁶ This can in turn have significant impact on the host and affect behavior, visceral sensitivity and inflammatory susceptibility.²⁵⁷⁻²⁶¹

Spinal Cord Plasticity and Glia Activation: Central Processing of Peripheral Pain Perception

Once peripheral sensitization has occurred, it activates the release of mediators in the spinal cord including growth factors^{262,263} (nerve growth factor or brain-derived neurotrophic factor) and upregulates some key ion channels and receptors such as acid-sensing ion channels 1A and neurokinin 1 receptor²⁶⁴⁻²⁶⁷ contributing to the phenomenon of spinal sensitization which has been associated with visceral hypersensitivity.

Very recently, spinal cord glia activation has been suggested as being another potential mechanism through which spinal sen-

sitization may occur in response to stress linked to the development and maintenance of visceral hypersensitivity.²⁶⁸⁻²⁷⁰ Candidate molecules involved in glia activation signaling include neurotransmitters such as substance P or glutamate, but also purinergic agents, opioids, chemokines and glucocorticoids (for review see reference²⁶⁸). Glutamate uptake through spinal glutamate transporters is critical for maintaining normal sensory transmission under physiologic conditions.^{271,272} A potential deficiency in glutamate reuptake by astrocytes associated with the activation of spinal cord glia²⁷³ has been recently suggested to play a role in the spinal sensitization and the development of visceral hypersensitivity in rats.²⁷⁴ Together, these data strongly support the concept that transmission of visceral nociceptive signals may be enhanced in various conditions of spinal microglia activation.²⁷⁵

Supraspinal Pain Modulation: A Fine-tuning between Pain Facilitation and Inhibition

Various supraspinal sites are involved in the modulation of visceral pain signals. Rectosigmoid distension in humans activates sensory (insula and somatosensory cortex), and limbic and paralimbic regions (including anterior cingulate cortex, amygdala and locus coeruleus).²⁷⁶ Many of these brain regions were also found to be significantly activated by CRD in rats.^{25-27,33,277}

The anterior cingulate cortex mediates key emotional-aversive aspects of pain and may also have a mnemonic role in which it allows transient storage of information during pain processing.^{189,278} Wistar-Kyoto rats, high-anxiety rats exhibiting visceral hypersensitivity¹¹¹ have greater prefrontal cortex activation in response to CRD compared to Sprague-Dawley.⁹¹ Another key limbic system structure that has been implicated in the affective component of pain is the central amygdala. It is involved in the processing of visceral information, attention, emotion and integrating the physical and psychological components of the stress response.²⁷⁹ It has also been found to contribute to visceral hypersensitivity in rats.²⁸⁰⁻²⁸³ Of relevance in the context of stress response, the CRF gene expression in the amygdaloid nucleus is upregulated in a mouse model of visceral pain and such a response is attenuated under conditions of anesthesia.^{283,284} Likewise, the locus coeruleus is a well established target of stress that expresses CRF₁ receptors, receives CRF innervation from nearby Barrington nucleus and increases firing in response to CRD that is mediated by CRF₁ receptor activation as shown by the use of CRF receptor antagonists and the responsiveness of LC neurons to both CRD and to central injection of CRF.^{53,285-290} Therefore these limbic and pontine sites are well positioned to co-

ordinate gut-brain interaction with visceral information from the gut impacting on cortical and limbic activities under conditions of stress-CRF₁ signaling activation which may modulate the visceral pain responses.^{60,76,291}

Thalamic relay nuclei have a key role in gating, filtering and processing sensory information en route to the cerebral cortex and are subject to similar activity-induced plasticity processes as the spinal cord.²⁹²⁻²⁹⁴ Upregulation of CRF₁ receptor in the thalamus is associated with visceral hyperalgesia in the rat model of neonatal maternal separation stress.²⁷⁵ Lastly, spinal visceral nociceptive reflexes are subject to facilitatory modulation from the rostroventral medulla, providing the basis for a mechanism by which visceral sensations can be enhanced from supraspinal sites^{295,296} under stress conditions associated with development of visceral hyperalgesia.²⁹⁷ Compromised engagement of descending pain inhibitory pathways as observed in maternally-stressed rats may also contribute to increase the visceral pain responses in those animals.¹¹⁷

Therapeutic Implications-Treatment Targeting Stress Reduction in Irritable Bowel Syndrome

The modulatory role of stress-related brain-gut interactions in the IBS pathophysiology, in particular neuroimmune modulation associated with psychological factors and emotional state^{16,189} has been confirmed by the encouraging outcome of non-pharmacologic and pharmacologic treatment modalities aimed at reducing stress perception.²⁹⁸⁻³⁰⁰ A broad range of evidence-based mind-body interventions including psychotherapy, cognitive behavioral therapy, hypnotherapy, relaxation exercises or mindfulness meditation has been shown to amend stress coping strategies, both at a cognitive level (catastrophic or self-defeating thoughts) and at a behavioral level (problem solving, especially interpersonal problems).^{300,301} The symptomatic improvement appears to result from the modulation of stress response, the autonomic nervous system balance restoration, and changes in the brain activation pattern in response to visceral stimuli. In addition to psychological mind-body approaches, clinical trials confirm the effectiveness of centrally-targeted pharmacological interventions such as with antidepressants, and anxiolytics, or combination of drugs from both groups in the treatment of chronic pain disorders.^{299,302,303} Many other pharmacological agents with anxiolytic and/or antidepressant properties, such as serotonergic and opioidergic agents, cannabinoid receptor 1 (CB₁) and somatosta-

tin receptors agonists, CRF₁, tachykinin and cholecystokinin receptors antagonists, have been recently shown to modulate stress-induced visceral hyperalgesia in animal models (for detailed review see reference³⁰⁴). Preliminary data suggest that anxiolytic activity of γ -aminobutyric acid-ergic agents (gabapentin) and $\alpha 2\delta$ ligand (pregabalin) may be also efficient in reducing central sensitization in hyperalgesia in clinical setting³⁰⁵ as shown in experimental models.³⁰⁶ New centrally acting agents providing analgesic effects include dexetofisopam (2,3-benzodiazepine receptor modulator) and quetiapine (atypical antipsychotic agent).³⁰⁷

Recent developments showing the critical interdependence between the composition and stability of the microbiota and GI sensory-motor function indicate a novel approach to IBS treatment with a use of probiotics, prebiotics and antibiotics.^{260,308} Specific modulation of the enteric microbiota in the context of neuroimmune interactions within the brain-gut axis opens a new promising strategy for stress-related disorders, particularly in the aspects of comorbidity in functional GI disorders such as IBS.²⁵⁷

However, some of the encouraging data from animal models concerning efficiency in alleviating stress-induced visceral hypersensitivity of such agents as CRF₁ receptor antagonist,³⁰⁹ CB₁/CB₂ receptor antagonist³¹⁰ or somatostatin receptor agonist (octreotide),³¹¹ are yet to be confirmed in clinical trials, especially with regard to global symptoms improvement and well-being. For example, CRF₁ receptor antagonists are being investigated in Phase II/III clinical trials for depression, anxiety and IBS.⁴² In fact, a recent clinical trial confirmed CRF₁ receptor antagonist efficacy in an anxiety model in healthy participants (7.5% CO₂ model).³¹² Some observed discrepancies between preclinical models and clinical trials may result from limited correlation between readout from animal studies being based on pseudoaffective reflex responses or unlearned behaviors and symptoms in IBS patients reflecting subjective pain experience highly modulated by cortical influences.¹ As discussed in this review, the methods used to monitor visceral sensitivity in rodents by inducing some bias in the observed responses could also potentially contribute to the lack of clinical translation of some drugs.

Amelioration of animal models of visceral pain, in their construct and face validity, particularly through the development of non-invasive methods to monitor visceral sensitivity together with a recently emerging algorithm of drug screening based on pharmacological brain imaging techniques opens promising venues in establishing an adequate approach in identifying effective treatment for IBS symptoms as well as IBS-related quality of life

impairment.

Acknowledgements

The authors thank Miss E. Hu for reviewing the manuscript.

References

1. Mayer EA, Bradesi S, Chang L, Spiegel BM, Bueller JA, Naliboff BD. Functional GI disorders: from animal models to drug development. *Gut* 2008;57:384-404.
2. Posserud I, Agerforz P, Ekman R, Björnsson ES, Abrahamsson H, Simrén M. Altered visceral perceptual and neuroendocrine response in patients with irritable bowel syndrome during mental stress. *Gut* 2004;53:1102-1108.
3. Elsenbruch S, Rosenberger C, Bingel U, Forsting M, Schedlowski M, Gizewski ER. Patients with irritable bowel syndrome have altered emotional modulation of neural responses to visceral stimuli. *Gastroenterology* 2010;139:1310-1319.
4. Elsenbruch S, Rosenberger C, Enck P, Forsting M, Schedlowski M, Gizewski ER. Affective disturbances modulate the neural processing of visceral pain stimuli in irritable bowel syndrome: an fMRI study. *Gut* 2010;59:489-495.
5. Lackner JM, Brasel AM, Quigley BM, et al. The ties that bind: perceived social support, stress, and IBS in severely affected patients. *Neurogastroenterol Motil* 2010;22:893-900.
6. Choung RS, Locke GR 3rd, Zinsmeister AR, Schleck CD, Talley NJ. Psychosocial distress and somatic symptoms in community subjects with irritable bowel syndrome: a psychological component is the rule. *Am J Gastroenterol* 2009;104:1772-1779.
7. Shen L, Kong H, Hou X. Prevalence of irritable bowel syndrome and its relationship with psychological stress status in Chinese university students. *J Gastroenterol Hepatol* 2009;24:1885-1890.
8. Blanchard EB, Lackner JM, Jaccard J, et al. The role of stress in symptom exacerbation among IBS patients. *J Psychosom Res* 2008;64:119-128.
9. Mayer EA, Naliboff BD, Chang L, Coutinho SV. V. Stress and irritable bowel syndrome. *Am J Physiol Gastrointest Liver Physiol* 2001;280:G519-G524.
10. Dufton LM, Konik B, Colletti R, et al. Effects of stress on pain threshold and tolerance in children with recurrent abdominal pain. *Pain* 2008;136:38-43.
11. Chitkara DK, van Tilburg MA, Blois-Martin N, Whitehead WE. Early life risk factors that contribute to irritable bowel syndrome in adults: a systematic review. *Am J Gastroenterol* 2008;103:765-774.
12. Videlock EJ, Adeyemo M, Licudine A, et al. Childhood trauma is associated with hypothalamic-pituitary-adrenal axis responsiveness in irritable bowel syndrome. *Gastroenterology* 2009;137:1954-1962.
13. Leserman J, Drossman DA. Relationship of abuse history to functional gastrointestinal disorders and symptoms: some possible mediating mechanisms. *Trauma Violence Abuse* 2007;8:331-343.
14. Gwee KA, Leong YL, Graham C, et al. The role of psychological

- and biological factors in postinfective gut dysfunction. *Gut* 1999;44: 400-406.
15. Spiller R, Garsed K. Infection, inflammation, and the irritable bowel syndrome. *Dig Liver Dis* 2009;41:844-849.
16. Mulak A, Bonaz B. Irritable bowel syndrome: a model of the brain-gut interactions. *Med Sci Monit* 2004;10:RA55-RA62.
17. Barreau F, Ferrier L, Fioramonti J, Bueno L. New insights in the etiology and pathophysiology of irritable bowel syndrome: contribution of neonatal stress models. *Pediatr Res* 2007;62:240-245.
18. Qin HY, Wu JC, Tong XD, Sung JJ, Xu HX, Bian ZX. Systematic review of animal models of post-infectious/post-inflammatory irritable bowel syndrome. *J Gastroenterol* 2011;46: 164-174.
19. Larauche M, Gourcerol G, Wang L, et al. Cortagine, a CRF1 agonist, induces stresslike alterations of colonic function and visceral hypersensitivity in rodents primarily through peripheral pathways. *Am J Physiol Gastrointest Liver Physiol* 2009;297:G215-G227.
20. Yarushkina NI. The role of hypothalamo-hypophyseal-adrenocortical system hormones in controlling pain sensitivity. *Neurosci Behav Physiol* 2008;38:759-766.
21. Coffin B, Bouhassira D, Sabate JM, Barbe L, Jian R. Alteration of the spinal modulation of nociceptive processing in patients with irritable bowel syndrome. *Gut* 2004;53:1465-1470.
22. Naliboff BD, Mayer EA. Brain imaging in IBS: drawing the line between cognitive and non-cognitive processes. *Gastroenterology* 2006;130:267-270.
23. Mayer EA, Naliboff BD, Craig AD. Neuroimaging of the brain-gut axis: from basic understanding to treatment of functional GI disorders. *Gastroenterology* 2006;131:1925-1942.
24. Lantéri-Minet M, Isnardon P, de PJ, Menétrey D. Spinal and hindbrain structures involved in viscerosception and visceronociception as revealed by the expression of Fos, Jun and Krox-24 proteins. *Neuroscience* 1993;55:737-753.
25. Wang L, Martinez V, Larauche M, Taché Y. Proximal colon distension induces Fos expression in oxytocin-, vasopressin-, CRF- and catecholamines-containing neurons in rat brain. *Brain Res* 2009;1247:79-91.
26. Martinez V, Wang L, Taché Y. Proximal colon distension induces Fos expression in the brain and inhibits gastric emptying through capsaicin-sensitive pathways in conscious rats. *Brain Res* 2006; 1086:168-180.
27. Mönnikes H, Rüter J, König M, et al. Differential induction of c-fos expression in brain nuclei by noxious and non-noxious colonic distension: role of afferent C-fibers and 5-HT₃ receptors. *Brain Res* 2003;966:253-264.
28. Murphy AZ, Suckow SK, Johns M, Traub RJ. Sex differences in the activation of the spinoparabrachial circuit by visceral pain. *Physiol Behav* 2009;97:205-212.
29. Wu JC, Ziea ET, LAo L, et al. Effect of electroacupuncture on visceral hyperalgesia, serotonin and fos expression in an animal model of irritable bowel syndrome. *J Neurogastroenterol Motil* 2010;16: 306-314.
30. Stam R, Ekkelenkamp K, Frankhuijzen AC, Bruijnzeel AW, Akkermans LM, Wiegant VM. Long-lasting changes in central nervous system responsivity to colonic distention after stress in rats. *Gastroenterology* 2002;123:1216-1225.
31. Traub RJ, Silva E, Gebhart GF, Solodkin A. Noxious colorectal distention induced-c-Fos protein in limbic brain structures in the rat. *Neurosci Lett* 1996;215:165-168.
32. Lazovic J, Wozos HF, Yang QX, et al. Regional activation in the rat brain during visceral stimulation detected by c-fos expression and fMRI. *Neurogastroenterol Motil* 2005;17:548-556.
33. Wang Z, Bradesi S, Maarek JM, et al. Regional brain activation in conscious, unrestrained rats in response to noxious visceral stimulation. *Pain* 2008;138:233-243.
34. Selye H. A syndrome produced by diverse noxious agents. *Nature* 1936;138:32.
35. Cannon WB. Bodily Changes in Pain, Hunger, Fear and Rage: an account of recent researches into the function of emotional excitement. New York and London: D. Appleton and company 1915:311.
36. Chrousos GP. Stress and disorders of the stress system. *Nat Rev Endocrinol* 2009;5:374-381.
37. McEwen BS. Stress, adaptation, and disease. Allostasis and allostatic load. *Ann N Y Acad Sci* 1998;840:33-44.
38. Sterling P, Eyer J. Allostasis: a new paradigm to explain arousal pathology. In: Fisher S, ed. *Handbook of life stress, cognition and health*. Oxford: John Wiley & Sons 1988:629-649.
39. McEwen BS, Stellar E. Stress and the individual. Mechanisms leading to disease. *Arch Intern Med* 1993;153:2093-2101.
40. Stengel A, Taché Y. Corticotropin-releasing factor signaling and visceral response to stress. *Exp Biol Med (Maywood)* 2010;235: 1168-1178.
41. Hauger RL, Grigoriadis DE, Dallman MF, Plotsky PM, Vale WW, Dautzenberg FM. International Union of Pharmacology. XXXVI. Current status of the nomenclature for receptors for corticotropin-releasing factor and their ligands. *Pharmacol Rev* 2003;55: 21-26.
42. Zorrilla EP, Koob GF. Progress in corticotropin-releasing factor-1 antagonist development. *Drug Discov Today* 2010;15:371-383.
43. Rivier CL, Grigoriadis DE, Rivier JE. Role of corticotropin-releasing factor receptors type 1 and 2 in modulating the rat adrenocorticotropin response to stressors. *Endocrinology* 2003;144:2396-2403.
44. Koob GF, Heinrichs SC. A role for corticotropin releasing factor and urocortin in behavioral responses to stressors. *Brain Res* 1999; 848:141-152.
45. Bale TL, Vale WW. CRF and CRF receptors: role in stress responsivity and other behaviors. *Annu Rev Pharmacol Toxicol* 2004;44:525-557.
46. Turnbull AV, Rivier C. Corticotropin-releasing factor (CRF) and endocrine responses to stress: CRF receptors, binding protein, and related peptides. *Proc Soc Exp Biol Med* 1997;215:1-10.
47. Taché Y, Martinez V, Million M, Wang L. Stress and the gastrointestinal tract III. Stress-related alterations of gut motor function: role of brain corticotropin-releasing factor receptors. *Am J Physiol Gastrointest Liver Physiol* 2001;280:G173-G177.
48. Caso JR, Leza JC, Menchen L. The effects of physical and psychological stress on the gastro-intestinal tract: lessons from animal models. *Curr Mol Med* 2008;8:299-312.
49. Friedman EM, Irwin MR. A role for CRH and the sympathetic nervous system in stress-induced immunosuppression. *Ann N Y*

- Acad Sci 1995;771:396-418.
50. Yorimitsu M, Okada S, Yamaguchi-Shima N, Shimizu T, Arai J, Yokotani K. Role of brain adrenoceptors in the corticotropin-releasing factor-induced central activation of sympatho-adrenomedullary outflow in rats. *Life Sci* 2008;82:487-494.
 51. Usui D, Yamaguchi-Shima N, Okada S, Shimizu T, Wakiguchi H, Yokotani K. Selective activation of the sympathetic ganglia by centrally administered corticotropin-releasing factor in rats. *Auton Neurosci* 2009;146:111-114.
 52. Tsatsanis C, Dermitzaki E, Venihaki M, et al. The corticotropin-releasing factor (CRF) family of peptides as local modulators of adrenal function. *Cell Mol Life Sci* 2007;64:1638-1655.
 53. Valentino RJ, Foote SL, Page ME. The locus coeruleus as a site for integrating corticotropin-releasing factor and noradrenergic mediation of stress responses. *Ann N Y Acad Sci* 1993;697:173-188.
 54. Kosyan HP, Wei JY, Taché Y. Intracisternal sauvagine is more potent than corticotropin-releasing factor to decrease gastric vagal efferent activity in rats. *Peptides* 1999;20:851-858.
 55. Wiersma A, Bohus B, Koolhaas JM. Corticotropin-releasing hormone microinfusion in the central amygdala diminishes a cardiac parasympathetic outflow under stress-free conditions. *Brain Res* 1993;625:219-227.
 56. Friedman EM, Irwin MR. Modulation of immune cell function by the autonomic nervous system. *Pharmacol Ther* 1997;74:27-38.
 57. Taché Y. The parasympathetic nervous system in the pathophysiology of the gastrointestinal tract. In: Bolis CL, Licinio J, Govoni S, eds. *Handbook of autonomic nervous system in health and diseases*. Chapter 15. New York: Marcel Dekker, Inc. 2002;463-503.
 58. Holsboer F, Ising M. Central CRH system in depression and anxiety - evidence from clinical studies with CRH1 receptor antagonists. *Eur J Pharmacol* 2008;583:350-357.
 59. Fukudo S. Role of corticotropin-releasing hormone in irritable bowel syndrome and intestinal inflammation. *J Gastroenterol* 2007;42(suppl 17):48-51.
 60. Taché Y, Brunnhuber S. From Hans Selye's discovery of biological stress to the identification of corticotropin-releasing factor signaling pathways: implication in stress-related functional bowel diseases. *Ann N Y Acad Sci* 2008;1148:29-41.
 61. Million M, Maillot C, Adelson DA, et al. Peripheral injection of sauvagine prevents repeated colorectal distension-induced visceral pain in female rats. *Peptides* 2005;26:1188-1195.
 62. Million M, Wang L, Wang Y, et al. CRF2 receptor activation prevents colorectal distension induced visceral pain and spinal ERK1/2 phosphorylation in rats. *Gut* 2006;55:172-181.
 63. Skórzewska A, Lehner M, Hamed A, et al. The effect of CRF2 receptor antagonists on rat conditioned fear responses and c-Fos and CRF expression in the brain limbic structures. *Behav Brain Res* 2011;221:155-165.
 64. Larauche M, Kiank C, Taché Y. Corticotropin releasing factor signaling in colon and ileum: regulation by stress and pathophysiological implications. *J Physiol Pharmacol* 2009;60(suppl 7):33-46.
 65. Basbaum AI, Bautista DM, Scherrer G, Julius D. Cellular and molecular mechanisms of pain. *Cell* 2009;139:267-284.
 66. Sengupta JN. Visceral pain: the neurophysiological mechanism. *Handb Exp Pharmacol* 2009;194:31-74.
 67. Robinson DR, Gebhart GF. Inside information: the unique features of visceral sensation. *Mol Interv* 2008;8:242-253.
 68. Grundy D. Neuroanatomy of visceral nociception: vagal and splanchnic afferent. *Gut* 2002;51(suppl 1):i2-i5.
 69. Blackshaw LA, Brookes SJ, Grundy D, Schemann M. Sensory transmission in the gastrointestinal tract. *Neurogastroenterol Motil* 2007;19:1-19.
 70. Ness TJ, Fillingim RB, Randich A, Backensto EM, Faught E. Low intensity vagal nerve stimulation lowers human thermal pain thresholds. *Pain* 2000;86:81-85.
 71. Randich A, Gebhart GF. Vagal afferent modulation of nociception. *Brain Res Brain Res Rev* 1992;17:77-99.
 72. Sugiura Y, Terui N, Hosoya Y, Tonosaki Y, Nishiyama K, Honda T. Quantitative analysis of central terminal projections of visceral and somatic unmyelinated (C) primary afferent fibers in the guinea pig. *J Comp Neurol* 1993;332:315-325.
 73. Wang G, Tang B, Traub RJ. Differential processing of noxious colonic input by thoracolumbar and lumbosacral dorsal horn neurons in the rat. *J Neurophysiol* 2005;94:3788-3794.
 74. Price DD. Central neural mechanisms that interrelate sensory and affective dimensions of pain. *Mol Interv* 2002;2:392-403, 339.
 75. Heinricher MM, Tavares I, Leith JL, Lumb BM. Descending control of nociception: Specificity, recruitment and plasticity. *Brain Res Rev* 2009;60:214-225.
 76. Tsuruoka M, Wang D, Tamaki J, Inoue T. Descending influence from the nucleus locus coeruleus/subcoeruleus on visceral nociceptive transmission in the rat spinal cord. *Neuroscience* 2010;165:1019-1024.
 77. Zhuo M, Gebhart GF. Facilitation and attenuation of a visceral nociceptive reflex from the rostroventral medulla in the rat. *Gastroenterology* 2002;122:1007-1019.
 78. Ness TJ, Gebhart GF. Colorectal distension as a noxious visceral stimulus: physiologic and pharmacologic characterization of pseudoaffective reflexes in the rat. *Brain Res* 1988;450:153-169.
 79. Christianson JA, Gebhart GF. Assessment of colon sensitivity by luminal distension in mice. *Nat Protoc* 2007;2:2624-2631.
 80. Larsson M, Arvidsson S, Ekman C, Bayati A. A model for chronic quantitative studies of colorectal sensitivity using balloon distension in conscious mice-effects of opioid receptor agonists. *Neurogastroenterol Motil* 2003;15:371-381.
 81. Bradesi S, Schwetz I, Ennes HS, et al. Repeated exposure to water avoidance stress in rats: a new model for sustained visceral hyperalgesia. *Am J Physiol Gastrointest Liver Physiol* 2005;289:G42-G53.
 82. Welting O, Van Den Wijngaard RM, De Jonge WJ, Holman R, Boeckxstaens GE. Assessment of visceral sensitivity using radio telemetry in a rat model of maternal separation. *Neurogastroenterol Motil* 2005;17:838-845.
 83. Nijssen MJ, Ongenae NG, Coulie B, Meulemans AL. Telemetric animal model to evaluate visceral pain in the freely moving rat. *Pain* 2003;105:115-123.
 84. Klueh U, Kreutzer DL. Murine model of implantable glucose sensors: a novel model for glucose sensor development. *Diabetes Technol Ther* 2005;7:727-737.
 85. Marois Y, Roy R, Vidovszky T, et al. Histopathological and immunological investigations of synthetic fibres and structures used in three prosthetic anterior cruciate ligaments: in vivo study in the rat.

- Biomaterials 1993;14:255-262.
86. Arvidsson S, Larsson M, Larsson H, Lindstrom E, Martinez V. Assessment of visceral pain-related pseudo-affective responses to colorectal distension in mice by intracolonic manometric recordings. *J Pain* 2006;7:108-118.
87. Tammperä A, Brusberg M, Axenborg J, Hirsch I, Larsson H, Lindström E. Evaluation of pseudo-affective responses to noxious colorectal distension in rats by manometric recordings. *Pain* 2005;116:220-226.
88. Larauche M, Gourcerol G, Million M, Adelson DW, Taché Y. Repeated psychological stress-induced alterations of visceral sensitivity and colonic motor functions in mice: Influence of surgery and postoperative single housing on visceromotor responses. *Stress* 2010;13:343-354.
89. Al-Chaer ED, Kawasaki M, Pasricha PJ. A new model of chronic visceral hypersensitivity in adult rats induced by colon irritation during postnatal development. *Gastroenterology* 2000;119:1276-1285.
90. Zhang XJ, Li Z, Chung EK, et al. Activation of extracellular signal-regulated protein kinase is associated with colorectal distension-induced spinal and supraspinal neuronal response and neonatal maternal separation-induced visceral hyperalgesia in rats. *J Mol Neurosci* 2009;37:274-287.
91. Gibney SM, Gosselin RD, Dinan TG, Cryan JF. Colorectal distension-induced prefrontal cortex activation in the Wistar-Kyoto rat: implications for irritable bowel syndrome. *Neuroscience* 2010;165:675-683.
92. Ait-Belgnaoui A, Eutamene H, Houdeau E, Bueno L, Fioramonti J, Theodorou V. *Lactobacillus farciminis* treatment attenuates stress-induced overexpression of Fos protein in spinal and supraspinal sites after colorectal distension in rats. *Neurogastroenterol Motil* 2009;21:567-569.
93. Johnson AC, Myers B, Lazovic J, Towner R, Greenwood-Van Meerveld B. Brain activation in response to visceral stimulation in rats with amygdala implants of corticosterone: an FMRI study. *PLoS One* 2010;5:e8573.
94. Coello C, Hjørnevik T, Courivaud F, Willoch F. Anatomical standardization of small animal brain FDG-PET images using synthetic functional template: Experimental comparison with anatomical template. *J Neurosci Methods* 2011;199:166-172.
95. Sawchenko PE, Li HY, Ericsson A. Circuits and mechanisms governing hypothalamic responses to stress: a tale of two paradigms. *Prog Brain Res* 2000;122:61-78.
96. Herman JP, Cullinan WE. Neurocircuitry of stress: central control of the hypothalamo-pituitary-adrenocortical axis. *Trends Neurosci* 1997;20:78-84.
97. Mayer EA, Collins SM. Evolving pathophysiologic models of functional gastrointestinal disorders. *Gastroenterology* 2002;122:2032-2048.
98. Rosenberger C, Elsenbruch S, Scholle A, et al. Effects of psychological stress on the cerebral processing of visceral stimuli in healthy women. *Neurogastroenterol Motil* 2009;21:740-e45.
99. Elsenbruch S. Abdominal pain in irritable bowel syndrome: a review of putative psychological, neural and neuro-immune mechanisms. *Brain Behav Immun* 2011;25:386-394.
100. Bouin M, Plourde V, Boivin M, et al. Rectal distention testing in patients with irritable bowel syndrome: sensitivity, specificity, and predictive values of pain sensory thresholds. *Gastroenterology* 2002;122:1771-1777.
101. Hotoleanu C, Popp R, Trifa AP, Nedelcu L, Dumitrascu DL. Genetic determination of irritable bowel syndrome. *World J Gastroenterol* 2008;14:6636-6640.
102. Camilleri M. Genetics and irritable bowel syndrome: from genomics to intermediate phenotype and pharmacogenetics. *Dig Dis Sci* 2009;54:2318-2324.
103. Zucchelli M, Camilleri M, Nixon Andreasson A, et al. Association of TNFSF15 polymorphism with irritable bowel syndrome. *Gut* Published Online First: 2 June 2011. doi:10.1136/gut.2011.241877
104. Markoutsaki T, Karantanos T, Gazouli M, Anagnou NP, Ladas SD, Karamanolis DG. Serotonin transporter and G protein beta 3 subunit gene polymorphisms in Greeks with irritable bowel syndrome. *Dig Dis Sci* Published Online First: 11 May 2011. doi: 10.1007/s10620-011-1726-7
105. Vazquez-Roque MI, Camilleri M, Carlson P, et al. HLA-DQ genotype is associated with accelerated small bowel transit in patients with diarrhea-predominant irritable bowel syndrome. *Eur J Gastroenterol Hepatol* 2011;23:481-487.
106. Dinan TG, Cryan J, Shanahan F, Keeling PW, Quigley EM. IBS: An epigenetic perspective. *Nat Rev Gastroenterol Hepatol* 2010;7:465-471.
107. Wu HH, Wang S. Strain differences in the chronic mild stress animal model of depression. *Behav Brain Res* 2010;213:94-102.
108. Porterfield VM, Zimomra ZR, Caldwell EA, Camp RM, Gabella KM, Johnson JD. Rat strain differences in restraint stress-induced brain cytokines. *Neuroscience* 2011;188:48-54.
109. O'Mahony CM, Clarke G, Gibney S, Dinan TG, Cryan JF. Strain differences in the neurochemical response to chronic restraint stress in the rat: relevance to depression. *Pharmacol Biochem Behav* 2011;97:690-699.
110. Shepard JD, Myers DA. Strain differences in anxiety-like behavior: association with corticotropin-releasing factor. *Behav Brain Res* 2008;186:239-245.
111. Gunter WD, Shepard JD, Foreman RD, Myers DA, Greenwood-Van Meerveld B. Evidence for visceral hypersensitivity in high-anxiety rats. *Physiol Behav* 2000;69:379-382.
112. Meaney MJ, Szyf M, Seckl JR. Epigenetic mechanisms of perinatal programming of hypothalamic-pituitary-adrenal function and health. *Trends Mol Med* 2007;13:269-277.
113. Sudo N, Chida Y, Aiba Y, et al. Postnatal microbial colonization programs the hypothalamic-pituitary-adrenal system for stress response in mice. *J Physiol* 2004;558:263-275.
114. O'Mahony SM, Hyland NP, Dinan TG, Cryan JF. Maternal separation as a model of brain-gut axis dysfunction. *Psychopharmacology (Berl)* 2011;214:71-88.
115. Rosztóczy A, Fioramonti J, Jarmay K, Barreau F, Wittmann T, Buéno L. Influence of sex and experimental protocol on the effect of maternal deprivation on rectal sensitivity to distension in the adult rat. *Neurogastroenterol Motil* 2003;15:679-686.
116. Plotsky PM, Meaney MJ. Early, postnatal experience alters hypothalamic corticotropin-releasing factor (CRF) mRNA, median eminence CRF content and stress-induced release in adult rats. *Brain Res Mol Brain Res* 1993;18:195-200.
117. Coutinho SV, Plotsky PM, Sablad M, et al. Neonatal maternal sep-

- aration alters stress-induced responses to viscerosomatic nociceptive stimuli in rat. *Am J Physiol Gastrointest Liver Physiol* 2002; 282:G307-G316.
118. Barreau F, Cartier C, Ferrier L, Fioramonti J, Bueno L. Nerve growth factor mediates alterations of colonic sensitivity and mucosal barrier induced by neonatal stress in rats. *Gastroenterology* 2004; 127:524-534.
 119. Lin C, Al-Chaer ED. Long-term sensitization of primary afferents in adult rats exposed to neonatal colon pain. *Brain Res* 2003; 971:73-82.
 120. Wang G, Ji Y, Lidow MS, Traub RJ. Neonatal hind paw injury alters processing of visceral and somatic nociceptive stimuli in the adult rat. *J Pain* 2004;5:440-449.
 121. LaPrairie JL, Murphy AZ. Long-term impact of neonatal injury in male and female rats: Sex differences, mechanisms and clinical implications. *Front Neuroendocrinol* 2010;31:193-202.
 122. LaPrairie JL, Murphy AZ. Neonatal injury alters adult pain sensitivity by increasing opioid tone in the periaqueductal gray. *Front Behav Neurosci* 2009;3:31.
 123. Bonaz B, Taché Y. Water-avoidance stress-induced c-fos expression in the rat brain and stimulation of fecal output: role of corticotropin-releasing factor. *Brain Res* 1994;641:21-28.
 124. Enck P, Merlin V, Erckenbrecht JF, Wienbeck M. Stress effects on gastrointestinal transit in the rat. *Gut* 1989;30:455-459.
 125. Eutamene H, Bradesi S, Larauche M, et al. Guanylate cyclase C-mediated antinociceptive effects of linacotide in rodent models of visceral pain. *Neurogastroenterol Motil* 2010;22:312-e84.
 126. Schwetz I, Bradesi S, McRoberts JA, et al. Delayed stress-induced colonic hypersensitivity in male Wistar rats: role of neurokinin-1 and corticotropin-releasing factor-1 receptors. *Am J Physiol Gastrointest Liver Physiol* 2004;286:G683-G691.
 127. Gué M, Del Rio-Lacheze C, Eutamene H, Théodorou V, Fioramonti J, Bueno L. Stress-induced visceral hypersensitivity to rectal distension in rats: role of CRF and mast cells. *Neurogastroenterol Motil* 1997;9:271-279.
 128. Chang L. Review article: epidemiology and quality of life in functional gastrointestinal disorders. *Aliment Pharmacol Ther* 2004; 20(suppl 7):31-39.
 129. Bennett EJ, Tennant CC, Piesse C, Badcock CA, Kellow JE. Level of chronic life stress predicts clinical outcome in irritable bowel syndrome. *Gut* 1998;43:256-261.
 130. Hong S, Fan J, Kemmerer ES, Evans S, Li Y, Wiley JW. Reciprocal changes in vanilloid (TRPV1) and endocannabinoid (CB1) receptors contribute to visceral hyperalgesia in the water avoidance stressed rat. *Gut* 2009;58:202-210.
 131. Larauche M, Bradesi S, Million M, et al. Corticotropin-releasing factor type 1 receptors mediate the visceral hyperalgesia induced by repeated psychological stress in rats. *Am J Physiol Gastrointest Liver Physiol* 2008;294:G1033-G1040.
 132. Larauche M, Mulak A, Kim YS, Million M, Taché Y. Sex differences in visceral sensitivity induced by repeated psychological stress in rats: differential role of opioid pathway. *Gut* 2010;59:A104.
 133. Larauche M, Mulak A, Yuan P-Q, Kanauchi O, Taché Y. Stress-induced visceral analgesia assessed non-invasively in rats is enhanced by prebiotic. *World J Gastroenterol* 2011 (In press)
 134. Green PG, Alvarez P, Gear RW, Mendoza D, Levine JD. Further validation of a model of fibromyalgia syndrome in the rat. *J Pain* Published Online First: 8 Apr 2011. doi:10.1016/j.jpain.2011.01.006
 135. Zheng J, Babygirija R, Bülbül M, Cerjak D, Ludwig K, Takahashi T. Hypothalamic oxytocin mediates adaptation mechanism against chronic stress in rats. *Am J Physiol Gastrointest Liver Physiol* 2010;299:G946-G953.
 136. Patel S, Hillard CJ. Adaptations in endocannabinoid signaling in response to repeated homotypic stress: a novel mechanism for stress habituation. *Eur J Neurosci* 2008;27:2821-2829.
 137. Winston JH, Xu GY, Sarna SK. Adrenergic stimulation mediates visceral hypersensitivity to colorectal distension following heterotypic chronic stress. *Gastroenterology* 2010;138:294-304.
 138. Savas LS, White DL, Wieman M, et al. Irritable bowel syndrome and dyspepsia among women veterans: prevalence and association with psychological distress. *Aliment Pharmacol Ther* 2009;29:115-125.
 139. White DL, Savas LS, Daci K, et al. Trauma history and risk of the irritable bowel syndrome in women veterans. *Aliment Pharmacol Ther* 2010;32:551-561.
 140. Cohen H, Jotkowitz A, Buskila D, et al. Post-traumatic stress disorder and other co-morbidities in a sample population of patients with irritable bowel syndrome. *Eur J Intern Med* 2006;17:567-571.
 141. Irwin C, Falsetti SA, Lydiard RB, Ballenger JC, Brock CD, Brenner W. Comorbidity of posttraumatic stress disorder and irritable bowel syndrome. *J Clin Psychiatry* 1996;57:576-578.
 142. Drossman DA, Leserman J, Nachman G, et al. Sexual and physical abuse in women with functional or organic gastrointestinal disorders. *Ann Intern Med* 1990;113:828-833.
 143. Klooker TK, Braak B, Painter RC, et al. Exposure to severe war-time conditions in early life is associated with an increased risk of irritable bowel syndrome: a population-based cohort study. *Am J Gastroenterol* 2009;104:2250-2256.
 144. Yehuda R, Schmeidler J, Labinsky E, et al. Ten-year follow-up study of PTSD diagnosis, symptom severity and psychosocial indices in aging holocaust survivors. *Acta Psychiatr Scand* 2009; 119:25-34.
 145. Stam R, Akkermans LM, Wiegant VM. Trauma and the gut: interactions between stressful experience and intestinal function. *Gut* 1997;40:704-709.
 146. Stam R. PTSD and stress sensitisation: a tale of brain and body Part 2: animal models. *Neurosci Biobehav Rev* 2007;31:558-584.
 147. Wang W, Liu Y, Zheng H, et al. A modified single-prolonged stress model for post-traumatic stress disorder. *Neurosci Lett* 2008;441:237-241.
 148. Rau V, DeCola JP, Fanselow MS. Stress-induced enhancement of fear learning: an animal model of posttraumatic stress disorder. *Neurosci Biobehav Rev* 2005;29:1207-1223.
 149. Stam R, van Laar TJ, Akkermans LM, Wiegant VM. Variability factors in the expression of stress-induced behavioural sensitisation. *Behav Brain Res* 2002;132:69-76.
 150. Collins SM, Vallance B, Barbara G, Borgaonkar M. Putative inflammatory and immunological mechanisms in functional bowel disorders. *Baillieres Best Pract Res Clin Gastroenterol* 1999;13: 429-436.
 151. Spiller RC. Postinfectious irritable bowel syndrome. *Gastroenterology* 2003;124:1662-1671.

152. Spiller R, Garsed K. Postinfectious irritable bowel syndrome. *Gastroenterology* 2009;136:1979-1988.
153. Long Y, Liu Y, Tong J, Qian W, Hou X. Effectiveness of trimebutine maleate on modulating intestinal hypercontractility in a mouse model of postinfectious irritable bowel syndrome. *Eur J Pharmacol* 2010;636:159-165.
154. Bercik P, Wang L, Verdú EF, et al. Visceral hyperalgesia and intestinal dysmotility in a mouse model of postinfective gut dysfunction. *Gastroenterology* 2004;127:179-187.
155. McLean PG, Picard C, Garcia-Villar R, Moré J, Fioramonti J, Buéno L. Effects of nematode infection on sensitivity to intestinal distension: role of tachykinin NK2 receptors. *Eur J Pharmacol* 1997;337:279-282.
156. Vergnolle N. Postinflammatory visceral sensitivity and pain mechanisms. *Neurogastroenterol Motil* 2008;20(suppl 1):73-80.
157. Keohane J, O'Mahony C, O'Mahony L, O'Mahony S, Quigley EM, Shanahan F. Irritable bowel syndrome-type symptoms in patients with inflammatory bowel disease: a real association or reflection of occult inflammation? *Am J Gastroenterol* 2010;105:1788, 1789-1794.
158. Long MD, Drossman DA. Inflammatory bowel disease, irritable bowel syndrome, or what?: a challenge to the functional-organic dichotomy. *Am J Gastroenterol* 2010;105:1796-1798.
159. Van Hoboken EA, Thijssen AY, Verhaaren R, et al. Symptoms in patients with ulcerative colitis in remission are associated with visceral hypersensitivity and mast cell activity. *Scand J Gastroenterol* Published Online First: 30 May 2011. doi:10.3109/00365521.2011.579156
160. Burton MB, Gebhart GF. Effects of intracolonic acetic acid on responses to colorectal distension in the rat. *Brain Res* 1995;672:77-82.
161. Palecek J, Willis WD. The dorsal column pathway facilitates visceromotor responses to colorectal distention after colon inflammation in rats. *Pain* 2003;104:501-507.
162. Ji Y, Tang B, Traub RJ. Modulatory effects of estrogen and progesterone on colorectal hyperalgesia in the rat. *Pain* 2005;117:433-442.
163. Traub RJ, Murphy A. Colonic inflammation induces fos expression in the thoracolumbar spinal cord increasing activity in the spinoparabrachial pathway. *Pain* 2002;95:93-102.
164. Coutinho SV, Meller ST, Gebhart GF. Intracolonic zymosan produces visceral hyperalgesia in the rat that is mediated by spinal NMDA and non-NMDA receptors. *Brain Res* 1996;736:7-15.
165. Gschossmann JM, Liebrechts T, Adam B, et al. Long-term effects of transient chemically induced colitis on the visceromotor response to mechanical colorectal distension. *Dig Dis Sci* 2004;49:96-101.
166. Adam B, Liebrechts T, Gschossmann JM, et al. Severity of mucosal inflammation as a predictor for alterations of visceral sensory function in a rat model. *Pain* 2006;123:179-186.
167. Zhou Q, Price DD, Caudle RM, Verne GN. Visceral and somatic hypersensitivity in a subset of rats following TNBS-induced colitis. *Pain* 2008;134:9-15.
168. Traub RJ, Tang B, Ji Y, Pandya S, Yfantis H, Sun Y. A rat model of chronic postinflammatory visceral pain induced by deoxycholic acid. *Gastroenterology* 2008;135:2075-2083.
169. Verma-Gandhu M, Verdú EF, Bercik P, et al. Visceral pain perception is determined by the duration of colitis and associated neuropeptide expression in the mouse. *Gut* 2007;56:358-364.
170. Larsson MH, Rapp L, Lindström E. Effect of DSS-induced colitis on visceral sensitivity to colorectal distension in mice. *Neurogastroenterol Motil* 2006;18:144-152.
171. Wlodarska M, Willing B, Keeney KM, et al. Antibiotic treatment alters the colonic mucus layer and predisposes the host to exacerbated *Citrobacter rodentium*-induced colitis. *Infect Immun* 2011;79:1536-1545.
172. Mendall MA, Kumar D. Antibiotic use, childhood affluence and irritable bowel syndrome (IBS). *Eur J Gastroenterol Hepatol* 1998;10:59-62.
173. Verdú EF, Bercik P, Verma-Gandhu M, et al. Specific probiotic therapy attenuates antibiotic induced visceral hypersensitivity in mice. *Gut* 2006;55:182-190.
174. Pimentel M, Chatterjee S, Chow EJ, Park S, Kong Y. Neomycin improves constipation-predominant irritable bowel syndrome in a fashion that is dependent on the presence of methane gas: sub-analysis of a double-blind randomized controlled study. *Dig Dis Sci* 2006;51:1297-1301.
175. Pimentel M, Lembo A, Chey WD, et al. Rifaximin therapy for patients with irritable bowel syndrome without constipation. *N Engl J Med* 2011;364:22-32.
176. Pimentel M, Morales W, Chua K, et al. Effects of Rifaximin Treatment and Retreatment in Nonconstipated IBS Subjects. *Dig Dis Sci* 2011;56:2067-2072.
177. Salonen A, de Vos WM, Palva A. Gastrointestinal microbiota in irritable bowel syndrome: present state and perspectives. *Microbiology* 2010;156(Pt 11):3205-3215.
178. Minocha A, Johnson WD, Wigginton WC. Prevalence of abdominal and pelvic surgeries in patients with irritable bowel syndrome: comparison between Caucasian and African Americans. *Am J Med Sci* 2008;335:82-88.
179. Li S, Yu Y, Prakash R. Possible pathogenetic roles of abdominal surgery in irritable bowel syndrome. *Med Hypotheses* 2011;76:497-499.
180. Miranda A, Peles S, Rudolph C, Shaker R, Sengupta JN. Altered visceral sensation in response to somatic pain in the rat. *Gastroenterology* 2004;126:1082-1089.
181. Cameron DM, Brennan TJ, Gebhart GF. Hind paw incision in the rat produces long-lasting colon hypersensitivity. *J Pain* 2008;9:246-253.
182. Whitehead WE, Palsson O, Jones KR. Systematic review of the comorbidity of irritable bowel syndrome with other disorders: what are the causes and implications? *Gastroenterology* 2002;122:1140-1156.
183. Pang X, Boucher W, Triadafilopoulos G, Sant GR, Theoharides TC. Mast cell and substance P-positive nerve involvement in a patient with both irritable bowel syndrome and interstitial cystitis. *Urology* 1996;47:436-438.
184. Peters KM, Killinger KA, Ibrahim IA. Childhood symptoms and events in women with interstitial cystitis/painful bladder syndrome. *Urology* 2009;73:258-262.
185. Pezzone MA, Liang R, Fraser MO. A model of neural cross-talk and irritation in the pelvis: implications for the overlap of chronic pelvic pain disorders. *Gastroenterology* 2005;128:1953-1964.
186. Miranda A, Mickle A, Schmidt J, et al. Neonatal cystitis-induced colonic hypersensitivity in adult rats: a model of viscerovisceral

- convergence. *Neurogastroenterol Motil* 2011;23:683-e281.
187. Van Oudenhove L, Coen SJ, Aziz Q. Functional brain imaging of gastrointestinal sensation in health and disease. *World J Gastroenterol* 2007;13:3438-3445.
 188. Seminowicz DA, Mikulis DJ, Davis KD. Cognitive modulation of pain-related brain responses depends on behavioral strategy. *Pain* 2004;112:48-58.
 189. Mayer EA, Tillisch K. The brain-gut axis in abdominal pain syndromes. *Annu Rev Med* 2011;62:381-396.
 190. Jarrett ME, Burr RL, Cain KC, Hertig V, Weisman P, Heitkemper MM. Anxiety and depression are related to autonomic nervous system function in women with irritable bowel syndrome. *Dig Dis Sci* 2003;48:386-394.
 191. Tougas G. The autonomic nervous system in functional bowel disorders. *Gut* 2000;47(suppl 4):iv78-iv80; discussion iv 87.
 192. Keita AV, Söderholm JD. The intestinal barrier and its regulation by neuroimmune factors. *Neurogastroenterol Motil* 2010;22:718-733.
 193. Bravo JA, Dinan TG, Cryan JF. Alterations in the central CRF system of two different rat models of comorbid depression and functional gastrointestinal disorders. *Int J Neuropsychopharmacol* 2011;14:666-683.
 194. Lemos JC, Zhang G, Walsh T, et al. Stress-hyperresponsive WKY rats demonstrate depressed dorsal raphe neuronal excitability and dysregulated CRF-mediated responses. *Neuropsychopharmacology* 2011;36:721-734.
 195. Overstreet DH, Djuric V. A genetic rat model of cholinergic hypersensitivity: implications for chemical intolerance, chronic fatigue, and asthma. *Ann N Y Acad Sci* 2001;933:92-102.
 196. Overstreet DH. The Flinders sensitive line rats: a genetic animal model of depression. *Neurosci Biobehav Rev* 1993;17:51-68.
 197. Elsenbruch S, Wang L, Hollerbach S, Schedlowski M, Tougas G. Pseudo-affective visceromotor responses and HPA axis activation following colorectal distension in rats with increased cholinergic sensitivity. *Neurogastroenterol Motil* 2004;16:801-809.
 198. Trimble N, Johnson AC, Foster A, Greenwood-van Meerveld B. Corticotropin-releasing factor receptor 1-deficient mice show decreased anxiety and colonic sensitivity. *Neurogastroenterol Motil* 2007;19:754-760.
 199. Million M, Wang L, Stenzel-Poore MP, et al. Enhanced pelvic responses to stressors in female CRF-overexpressing mice. *Am J Physiol Regul Integr Comp Physiol* 2007;292:R1429-R1438.
 200. Stenzel-Poore MP, Cameron VA, Vaughan J, Sawchenko PE, Vale W. Development of Cushing's syndrome in corticotropin-releasing factor transgenic mice. *Endocrinology* 1992;130:3378-3386.
 201. Kimura M, Müller-Preuss P, Lu A, et al. Conditional corticotropin-releasing hormone overexpression in the mouse forebrain enhances rapid eye movement sleep. *Mol Psychiatry* 2010;15:154-165.
 202. Lu A, Steiner MA, Whittle N, et al. Conditional mouse mutants highlight mechanisms of corticotropin-releasing hormone effects on stress-coping behavior. *Mol Psychiatry* 2008;13:1028-1042.
 203. Bakshi VP, Kalin NH. Corticotropin-releasing hormone and animal models of anxiety: gene-environment interactions. *Biol Psychiatry* 2000;48:1175-1198.
 204. Kolber BJ, Bovle MP, Wiczorek, et al. Transient early-life forebrain corticotropin-releasing hormone elevation causes long-lasting anxiogenic and despair-like changes in mice. *J Neurosci* 2010;30:2571-2581.
 205. Deussing JM, Wurst W. Dissecting the genetic effect of the CRH system on anxiety and stress-related behaviour. *C R Biol* 2005;328:199-212.
 206. Delic S, Streif S, Deussing JM, et al. Genetic mouse models for behavioral analysis through transgenic RNAi technology. *Genes Brain Behav* 2008;7:821-830.
 207. Butler RK, Finn DP. Stress-induced analgesia. *Prog Neurobiol* 2009;88:184-202.
 208. Gui X, Carraway RE, Dobner PR. Endogenous neurotensin facilitates visceral nociception and is required for stress-induced antinociception in mice and rats. *Neuroscience* 2004;126:1023-1032.
 209. Schwetz I, McRoberts JA, Coutinho SV, et al. Corticotropin-releasing factor receptor 1 mediates acute and delayed stress-induced visceral hyperalgesia in maternally separated Long-Evans rats. *Am J Physiol Gastrointest Liver Physiol* 2005;289:G704-G712.
 210. Rivat C, Laboureyras E, Laulin JP, Le RC, Richebe P, Simonnet G. Non-nociceptive environmental stress induces hyperalgesia, not analgesia, in pain and opioid-experienced rats. *Neuropsychopharmacology* 2007;32:2217-2228.
 211. Berman SM, Naliboff BD, Suyenobu B, et al. Reduced brainstem inhibition during anticipated pelvic visceral pain correlates with enhanced brain response to the visceral stimulus in women with irritable bowel syndrome. *J Neurosci* 2008;28:349-359.
 212. Piche M, Arsenault M, Poitras P, Rainville P, Bouin M. Widespread hypersensitivity is related to altered pain inhibition processes in irritable bowel syndrome. *Pain* 2010;148:49-58.
 213. Song GH, Venkatraman V, Ho KY, Chee MW, Yeoh KG, Wilder-Smith CH. Cortical effects of anticipation and endogenous modulation of visceral pain assessed by functional brain MRI in irritable bowel syndrome patients and healthy controls. *Pain* 2006;126:79-90.
 214. Wilder-Smith CH, Schindler D, Lovblad K, Redmond SM, Nirkko A. Brain functional magnetic resonance imaging of rectal pain and activation of endogenous inhibitory mechanisms in irritable bowel syndrome patient subgroups and healthy controls. *Gut* 2004;53:1595-1601.
 215. Heitkemper MM, Chang L. Do fluctuations in ovarian hormones affect gastrointestinal symptoms in women with irritable bowel syndrome? *Gend Med* 2009;6(suppl 2):152-167.
 216. Adeyemo MA, Spiegel BM, Chang L. Meta-analysis: do irritable bowel syndrome symptoms vary between men and women? *Aliment Pharmacol Ther* 2010;32:738-755.
 217. Fillingim RB, King CD, Ribeiro-Dasilva MC, Rahim-Williams B, Riley JL 3rd. Sex, gender, and pain: a review of recent clinical and experimental findings. *J Pain* 2009;10:447-485.
 218. Mulak A, Taché Y. Sex difference in irritable bowel syndrome: do gonadal hormones play a role? *Gastroenterol Pol* 2010;17:89-97.
 219. Koch KM, Palmer JL, Noordin N, Tomlinson JJ, Baidoo C. Sex and age differences in the pharmacokinetics of alosetron. *Br J Clin Pharmacol* 2002;53:238-242.
 220. Taché Y, Million M, Nelson AG, Lamy C, Wang L. Role of corticotropin-releasing factor pathways in stress-related alterations of colonic motor function and viscerosensitivity in female rodents. *Gend Med* 2005;2:146-154.

221. Aloisi AM, Affaitati G, Ceccarelli I, et al. Estradiol and testosterone differently affect visceral pain-related behavioural responses in male and female rats. *Eur J Pain* 2010;14:602-607.
222. Ji Y, Tang B, Traub RJ. The visceromotor response to colorectal distention fluctuates with the estrous cycle in rats. *Neuroscience* 2008;154:1562-1567.
223. Holdcroft A, Sapsed-Byrne S, Ma D, Hammal D, Forsling ML. Sex and oestrous cycle differences in visceromotor responses and vasopressin release in response to colonic distension in male and female rats anaesthetized with halothane. *Br J Anaesth* 2000;85:907-910.
224. Sapsed-Byrne S, Ma D, Ridout D, Holdcroft A. Estrous cycle phase variations in visceromotor and cardiovascular responses to colonic distension in the anesthetized rat. *Brain Res* 1996;742:10-16.
225. Ouyang A, Wrzros HF. Contribution of gender to pathophysiology and clinical presentation of IBS: should management be different in women? *Am J Gastroenterol* 2006;101(suppl 12):S602-S609.
226. Azpiroz F, Bouin M, Camilleri M, et al. Mechanisms of hypersensitivity in IBS and functional disorders. *Neurogastroenterol Motil* 2007;19(suppl 1):62-88.
227. Lembo T, Plourde V, Shui Z, et al. Effects of the corticotropin-releasing factor (CRF) on rectal afferent nerves in humans. *Neurogastroenterol Motil* 1996;8:9-18.
228. Nozu T, Kudaira M. Corticotropin-releasing factor induces rectal hypersensitivity after repetitive painful rectal distention in healthy humans. *J Gastroenterol* 2006;41:740-744.
229. Sagami Y, Shimada Y, Tayama J, et al. Effect of a corticotropin releasing hormone receptor antagonist on colonic sensory and motor function in patients with irritable bowel syndrome. *Gut* 2004;53:958-964.
230. Tayama J, Sagami Y, Shimada Y, Hongo M, Fukudo S. Effect of alpha-helical CRH on quantitative electroencephalogram in patients with irritable bowel syndrome. *Neurogastroenterol Motil* 2007;19:471-483.
231. La JH, Sung TS, Kim HJ, Kim TW, Kang TM, Yang IS. Peripheral corticotropin releasing hormone mediates post-inflammatory visceral hypersensitivity in rats. *World J Gastroenterol* 2008;14:731-736.
232. Barreau F, Cartier C, Leveque M, et al. Pathways involved in gut mucosal barrier dysfunction induced in adult rats by maternal deprivation: corticotrophin-releasing factor and nerve growth factor interplay. *J Physiol* 2007;580(Pt 1):347-356.
233. Wallon C, Yang PC, Keita AV, et al. Corticotropin-releasing hormone (CRH) regulates macromolecular permeability via mast cells in normal human colonic biopsies in vitro. *Gut* 2008;57:50-58.
234. Barbara G, Wang B, Stanghellini V, et al. Mast cell-dependent excitation of visceral-nociceptive sensory neurons in irritable bowel syndrome. *Gastroenterology* 2007;132:26-37.
235. Van den Wijngaard RM, Klooker TK, de Jonge WJ, Boeckstaens GE. Peripheral relays in stress-induced activation of visceral afferents in the gut. *Auton Neurosci* 2010;153:99-105.
236. Cenac N, Andrews CN, Holzhausen M, et al. Role for protease activity in visceral pain in irritable bowel syndrome. *J Clin Invest* 2007;117:636-647.
237. Gold MS, Zhang L, Wrigley DL, Traub RJ. Prostaglandin E(2) modulates TTX-R I(Na) in rat colonic sensory neurons. *J Neurophysiol* 2002;88:1512-1522.
238. Van den Wijngaard RM, Kooker TK, Welting O, et al. Essential role for TRPV1 in stress-induced (mast cell-dependent) colonic hypersensitivity in maternally separated rats. *Neurogastroenterol Motil* 2009;21:1107-e94.
239. Holzer P. Gastrointestinal afferents as targets of novel drugs for the treatment of functional bowel disorders and visceral pain. *Eur J Pharmacol* 2001;429:177-193.
240. Jones RC 3rd, Xu L, Gebhart GF. The mechanosensitivity of mouse colon afferent fibers and their sensitization by inflammatory mediators require transient receptor potential vanilloid 1 and acid-sensing ion channel 3. *J Neurosci* 2005;25:10981-10989.
241. Cregg R, Momin A, Rugiero F, Wood JN, Zhao J. Pain channelopathies. *J Physiol* 2010;588(Pt 11):1897-1904.
242. McRoberts JA, Coutinho SV, Marvizón JC, et al. Role of peripheral N-methyl-D-aspartate (NMDA) receptors in visceral nociception in rats. *Gastroenterology* 2001;120:1737-1748.
243. Yu YB, Yang J, Zuo XL, Gao LJ, Wang P, Li YQ. Transient receptor potential vanilloid-1 (TRPV1) and ankyrin-1 (TRPA1) participate in visceral hyperalgesia in chronic water avoidance stress rat model. *Neurochem Res* 2010;35:797-803.
244. Winston J, Shenoy M, Medley D, Naniwadekar A, Pasricha PJ. The vanilloid receptor initiates and maintains colonic hypersensitivity induced by neonatal colon irritation in rats. *Gastroenterology* 2007;132:615-627.
245. Ravnefjord A, Brusberg M, Kang D, et al. Involvement of the transient receptor potential vanilloid 1 (TRPV1) in the development of acute visceral hyperalgesia during colorectal distension in rats. *Eur J Pharmacol* 2009;611:85-91.
246. Piche T, Barbara G, Aubert P, et al. Impaired intestinal barrier integrity in the colon of patients with irritable bowel syndrome: involvement of soluble mediators. *Gut* 2009;58:196-201.
247. Ait-Belgnaoui A, Bradesi S, Fioramonti J, Theodorou V, Bueno L. Acute stress-induced hypersensitivity to colonic distension depends upon increase in paracellular permeability: role of myosin light chain kinase. *Pain* 2005;113:141-147.
248. Zhou Q, Zhang B, Verne GN. Intestinal membrane permeability and hypersensitivity in the irritable bowel syndrome. *Pain* 2009;146:41-46.
249. Santos J, Yang PC, Söderholm JD, Benjamin M, Perdue MH. Role of mast cells in chronic stress induced colonic epithelial barrier dysfunction in the rat. *Gut* 2001;48:630-636.
250. Söderholm JD, Yang PC, Ceponis P, et al. Chronic stress induces mast cell-dependent bacterial adherence and initiates mucosal inflammation in rat intestine. *Gastroenterology* 2002;123:1099-1108.
251. Yu LC, Perdue MH. Role of mast cells in intestinal mucosal function: studies in models of hypersensitivity and stress. *Immunol Rev* 2001;179:61-73.
252. Vicario M, Guilarte M, Alonso C, et al. Chronological assessment of mast cell-mediated gut dysfunction and mucosal inflammation in a rat model of chronic psychosocial stress. *Brain Behav Immun* 2010;24:1166-1175.
253. Demaude J, Salvador-Cartier C, Fioramonti J, Ferrier L, Bueno L. Phenotypic changes in colonocytes following acute stress or activation of mast cells in mice: implications for delayed epithelial barrier dysfunction. *Gut* 2006;55:655-661.

254. Bailey MT, Dowd SE, Galley JD, Hufnagle AR, Allen RG, Lyte M. Exposure to a social stressor alters the structure of the intestinal microbiota: Implications for stressor-induced immunomodulation. *Brain Behav Immun* 2011;25:397-407.
255. Bailey MT, Dowd SE, Parry NM, Galley JD, Schauer DB, Lyte M. Stressor exposure disrupts commensal microbial populations in the intestines and leads to increased colonization by *Citrobacter rodentium*. *Infect Immun* 2010;78:1509-1519.
256. O'Mahony SM, Marchesi JR, Scully P, et al. Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. *Biol Psychiatry* 2009;65:263-267.
257. Cryan JF, O'Mahony SM. The microbiome-gut-brain axis: from bowel to behavior. *Neurogastroenterol Motil* 2011;23:187-192.
258. Bercik P. The microbiota-gut-brain axis: learning from intestinal bacteria? *Gut* 2011;60:288-289.
259. Heijtz RD, Wang S, Anuar F, et al. Normal gut microbiota modulates brain development and behavior. *Proc Natl Acad Sci U S A* 2011;108:3047-3052.
260. Rhee SH, Pothoulakis C, Mayer EA. Principles and clinical implications of the brain-gut-enteric microbiota axis. *Nat Rev Gastroenterol Hepatol* 2009;6:306-314.
261. Collins SM, Bercik P. The relationship between intestinal microbiota and the central nervous system in normal gastrointestinal function and disease. *Gastroenterology* 2009;136:2003-2014.
262. Chung EK, Zhang XJ, Xu HX, Sung JJ, Bian ZX. Visceral hyperalgesia induced by neonatal maternal separation is associated with nerve growth factor-mediated central neuronal plasticity in rat spinal cord. *Neuroscience* 2007;149:685-695.
263. Chung EK, Bian ZX, Xu HX, Sung JJ. Neonatal maternal separation increases brain-derived neurotrophic factor and tyrosine kinase receptor B expression in the descending pain modulatory system. *Neurosignals* 2009;17:213-221.
264. Matricon J, Gelot A, Etienne M, Lazdunski M, Muller E, Ardid D. Spinal cord plasticity and acid-sensing ion channels involvement in a rodent model of irritable bowel syndrome. *Eur J Pain* 2011;15:335-343.
265. Gaudreau GA, Plourde V. Role of tachykinin NK1, NK2 and NK3 receptors in the modulation of visceral hypersensitivity in the rat. *Neurosci Lett* 2003;351:59-62.
266. Bradesi S, Kokkotou E, Simeonidis S, et al. The role of neurokinin 1 receptors in the maintenance of visceral hyperalgesia induced by repeated stress in rats. *Gastroenterology* 2006;130:1729-1742.
267. Bradesi S, Svensson CI, Steinauer J, Pothoulakis C, Yaksh TL, Mayer EA. Role of spinal microglia in visceral hyperalgesia and NK1R up-regulation in a rat model of chronic stress. *Gastroenterology* 2009;136:1339-1348, e1-e2.
268. Bradesi S. Role of spinal cord glia in the central processing of peripheral pain perception. *Neurogastroenterol Motil* 2010;22:499-511.
269. Saab CY, Park YC, Al-Chaer ED. Thalamic modulation of visceral nociceptive processing in adult rats with neonatal colon irritation. *Brain Res* 2004;1008:186-192.
270. Bradesi S, Svensson CI, Steinauer J, Pothoulakis C, Yaksh TL, Mayer EA. Role of spinal microglia activation in visceral hyperalgesia following chronic psychological stress in Wistar rats. *Gastroenterology* 2009;136:1339-1348.
271. Liaw WJ, Stephens RL Jr, Binns BC, et al. Spinal glutamate uptake is critical for maintaining normal sensory transmission in rat spinal cord. *Pain* 2005;115:60-70.
272. Lin Y, Tian G, Roman K, et al. Increased glial glutamate transporter EAAT2 expression reduces visceral nociceptive response in mice. *Am J Physiol Gastrointest Liver Physiol* 2009;296:G129-G134.
273. Svensson CI, Hua XY, Protter AA, Powell HC, Yaksh TL. Spinal p38 MAP kinase is necessary for NMDA-induced spinal PGE(2) release and thermal hyperalgesia. *Neuroreport* 2003;14:1153-1157.
274. Gosselin RD, O'Connor RM, Tramullas M, Julio-Pieper M, Dinan TG, Cryan JF. Riluzole normalizes early-life stress-induced visceral hypersensitivity in rats: role of spinal glutamate reuptake mechanisms. *Gastroenterology* 2010;138:2418-2425.
275. Tjong YW, Ip SP, Lao L, et al. Neonatal maternal separation elevates thalamic corticotrophin releasing factor type 1 receptor expression response to colonic distension in rat. *Neuro Endocrinol Lett* 2010;31:215-220.
276. Tillisch K, Mayer EA, Labus JS. Quantitative meta-analysis identifies brain regions activated during rectal distension in irritable bowel syndrome. *Gastroenterology* 2011;140:91-100.
277. Zhang R, Zou N, Li J, et al. Elevated expression of c-fos in central nervous system correlates with visceral hypersensitivity in irritable bowel syndrome (IBS): a new target for IBS treatment. *Int J Colorectal Dis* Published Online First: 22 Feb 2011. doi:10.1016/j.expneurol.2011.04.020
278. Chang L. Brain responses to visceral and somatic stimuli in irritable bowel syndrome: a central nervous system disorder? *Gastroenterol Clin North Am* 2005;34:271-279.
279. Dayas CV, Buller KM, Crane JW, Xu Y, Day TA. Stressor categorization: acute physical and psychological stressors elicit distinctive recruitment patterns in the amygdala and in medullary noradrenergic cell groups. *Eur J Neurosci* 2001;14:1143-1152.
280. Greenwood-Van Meerveld B, Gibson M, Gunter W, Shepard J, Foreman R, Myers D. Stereotaxic delivery of corticosterone to the amygdala modulates colonic sensitivity in rats. *Brain Res* 2001;893:135-142.
281. Myers B, Dittmeyer K, Greenwood-Van MB. Involvement of amygdaloid corticosterone in altered visceral and somatic sensation. *Behav Brain Res* 2007;181:163-167.
282. Myers B, Greenwood-Van Meerveld B. Corticosteroid receptor-mediated mechanisms in the amygdala regulate anxiety and colonic sensitivity. *Am J Physiol Gastrointest Liver Physiol* 2007;292:G1622-G1629.
283. Kim SH, Han JE, Hwang S, Oh DH. The expression of corticotropin-releasing factor in the central nucleus of the amygdala, induced by colorectal distension, is attenuated by general anesthesia. *J Korean Med Sci* 2010;25:1646-1651.
284. Nishii H, Nomura M, Aono H, Fujimoto N, Matsumoto T. Up-regulation of galanin and corticotropin-releasing hormone mRNAs in the key hypothalamic and amygdaloid nuclei in a mouse model of visceral pain. *Regul Pept* 2007;141:105-112.
285. Kosoyan HP, Grigoriadis DE, Taché Y. The CRF(1) receptor antagonist, NBI-35965, abolished the activation of locus coeruleus neurons induced by colorectal distension and intracisternal CRF in rats. *Brain Res* 2005;1056:85-96.

286. Curtis AL, Pavcovich LA, Grigoriadis DE, Valentino RJ. Previous stress alters corticotropin-releasing factor neurotransmission in the locus coeruleus. *Neuroscience* 1995;65:541-550.
287. Lechner SM, Curtis AL, Brons R, Valentino RJ. Locus coeruleus activation by colon distention: role of corticotropin-releasing factor and excitatory amino acids. *Brain Res* 1997;756:114-124.
288. Rouzade-Dominguez ML, Curtis AL, Valentino RJ. Role of Barrington's nucleus in the activation of rat locus coeruleus neurons by colonic distension. *Brain Res* 2001;917:206-218.
289. Reyes BA, Glaser JD, Van Bockstaele EJ. Ultrastructural evidence for co-localization of corticotropin-releasing factor receptor and mu-opioid receptor in the rat nucleus locus coeruleus. *Neurosci Lett* 2007;413:216-221.
290. Reyes BA, Valentino RJ, Van Bockstaele EJ. Stress-induced intracellular trafficking of corticotropin-releasing factor receptors in rat locus coeruleus neurons. *Endocrinology* 2008;149:122-130.
291. Valentino RJ, Miselis RR, Pavcovich LA. Pontine regulation of pelvic viscera: pharmacological target for pelvic visceral dysfunctions. *Trends Pharmacol Sci* 1999;20:253-260.
292. Kuner R. Central mechanisms of pathological pain. *Nat Med* 2010;16:1258-1266.
293. Cheong E, Lee S, Choi BJ, Sun M, Lee CJ, Shin HS. Tuning thalamic firing modes via simultaneous modulation of T- and L-type Ca^{2+} channels controls pain sensory gating in the thalamus. *J Neurosci* 2008;28:13331-13340.
294. Ren Y, Zhang L, Lu Y, Yang H, Westlund KN. Central lateral thalamic neurons receive noxious visceral mechanical and chemical input in rats. *J Neurophysiol* 2009;102:244-258.
295. Zhuo M, Gebhart GF. Facilitation and attenuation of a visceral nociceptive reflex from the rostroventral medulla in the rat. *Gastroenterology* 2002;122:1007-1019.
296. Sanoja R, Tortorici V, Fernandez C, Price TJ, Cervero F. Role of RVM neurons in capsaicin-evoked visceral nociception and referred hyperalgesia. *Eur J Pain* 2010;14:120.e1-e9.
297. Martenson ME, Cetas JS, Heinricher MM. A possible neural basis for stress-induced hyperalgesia. *Pain* 2009;142:236-244.
298. Kearney DJ, Brown-Chang J. Complementary and alternative medicine for IBS in adults: mind-body interventions. *Nat Clin Pract Gastroenterol Hepatol* 2008;5:624-636.
299. Palsson OS, Drossman DA. Psychiatric and psychological dysfunction in irritable bowel syndrome and the role of psychological treatments. *Gastroenterol Clin North Am* 2005;34:281-303.
300. Whorwell PJ. Behavioral therapy for IBS. *Nat Clin Pract Gastroenterol Hepatol* 2009;6:148-149.
301. Blanchard EB, Lackner JM, Sanders K, et al. A controlled evaluation of group cognitive therapy in the treatment of irritable bowel syndrome. *Behav Res Ther* 2007;45:633-648.
302. Warnock JK, Clayton AH. Chronic episodic disorders in women. *Psychiatr Clin North Am* 2003;26:725-740.
303. Verdu B, Decosterd I, Buclin T, Stiefel F, Berney A. Antidepressants for the treatment of chronic pain. *Drugs* 2008;68:2611-2632.
304. Larauche M, Mulak A, Taché Y. Stress and visceral pain: from animal models to clinical therapies. *Exp Neurol* Published Online First: 6 May 2011. doi:10.1016/j.expneurol.2011.04.020
305. Camilleri M, Andresen V. Current and novel therapeutic options for irritable bowel syndrome management. *Dig Liver Dis* 2009;41:854-862.
306. Million M, Wang L, Adelson DW, Roman F, Diop L, Taché Y. Pregabalin decreases visceral pain and prevents spinal neuronal activation in rats. *Gut* 2007;56:1482-1484.
307. Camilleri M. Review article: new receptor targets for medical therapy in irritable bowel syndrome. *Aliment Pharmacol Ther* 2010;31:35-46.
308. Collins SM, Denou E, Verdu EF, Bercik P. The putative role of the intestinal microbiota in the irritable bowel syndrome. *Dig Liver Dis* 2009;41:850-853.
309. Dukes GE, Mayer EA, Kelleher DL, Hicks KJ, Boardley RL, Alpers DH. A randomized, double blind, placebo (PLA) controlled, crossover study to evaluate the efficacy and safety of the corticotropin releasing factor 1 (CRF₁) receptor antagonist (RA) GW876008 in irritable bowel syndrome (IBS) patients (Pts). *Neurogastroenterol Motil* 2009;21(suppl 1):84.
310. Klooker TK, Leliefeld KE, van den Wijngaard RM, Boeckstaens GE. The cannabinoid receptor agonist delta-9-tetrahydrocannabinol does not affect visceral sensitivity to rectal distension in healthy volunteers and IBS patients. *Neurogastroenterol Motil* 2011;23:30-35, e2.
311. Klooker TK, Kuiken SD, Lei A, Boeckstaens GE. Effect of long-term treatment with octreotide on rectal sensitivity in patients with non-constipated irritable bowel syndrome. *Aliment Pharmacol Ther* 2007;26:605-615.
312. Bailey JE, Papadopoulos A, Diaper A, et al. Preliminary evidence of anxiolytic effects of the CRF1 receptor antagonist R317573 in the 7.5% CO₂ proof-of-concept experimental model of human anxiety. *J Psychopharmacol* Published Online First: 9 May 2011. doi:10.1177/0269881111400650